



Silk Fibroin in Bioelectronics Structural Engineering, Oriented Crystallization, And Long-Term Implantable Interface Strategies.

Harish N¹, Panneer Pandiyan Premkumar*, Jemimah smileyn J², Hariny Raja³, Srijitha G⁴, Vaishnavi M⁵, Avanthika E⁶, Pavithra R⁷, Praveen R⁸, Priyadharshnini M⁹, Subash Chandra Bose R¹⁰

¹Department of pharmacy, Saveetha college of pharmacy, SIMATS, Chennai, Tamilnadu, ²Department of pharmaceuticals, SIMATS College of pharmacy, Saveetha Institute Of Medical and Technical Sciences, Chennai, Tamilnadu, India

Corresponding Author

Panneer Pandian Premkumar

Professor, Department of Pharmaceutics, Saveetha College of Pharmacy, Saveetha Institute of Medical and Technical Sciences (SIMATS), Thandalam, Chennai, 602105, India.

(Received: 16 March 2026

Revised: 14 April 2026

Accepted: 01 May 2026)

KEYWORDS

Silk Fibroin (SF),
Bioelectronics,
Biomaterials,
Implantable Devices,
Wearable Sensors,
Neural Interfaces,
Biocompatibility,
Biodegradability,
Flexible Electronics.

ABSTRACT:

Introduction: Silk fibroin (SF) is an attractive biomaterial for bioelectronic devices because of its biocompatibility, mechanical and optical properties, and water processing capability. But traditional SF-based bioelectronic devices have poor long-term stability due to uncontrolled degradation, moisture-induced instability, biofouling and fatigue.

Objectives The objective of this review is to present recent progress in structural engineering methods such as oriented crystallization and pre-stretching, to enhance stability and functionality of SF-based bioelectronic devices.

Methods: The literature of modified SF materials was reviewed with methods of improving molecular orientation and β -sheet formation. Their effects on mechanical properties, dielectric responses, degradation rate and electrical conductivity were evaluated, as well as their in vivo stability and biocompatibility.

Results Modifications resulted in enhanced mechanical and dielectric stability, and controllable degradation. Crystallinity improved tunability of conductivity, thus ensuring electrical stability. Likewise, increased moisture and biofouling resistance and better in vivo compatibility highlight the potential of engineered SF for prolonged use in neurointerfaces, wearable sensors, and in vivo.

Conclusions: Structurally engineered silk fibroin shows great promise as a durable and dependable material for long-term bioelectronic applications. By improving molecular alignment and controlling crystallinity, many of the limitations seen in traditional silk-based systems can be effectively addressed. These advancements enhance stability and performance, making silk fibroin a strong candidate for use in long-term implantable devices, including neural interfaces, wearable sensors, and continuous physiological monitoring systems..

1.0 Introduction Silk Fibroin in Modern Bioelectronics

Silk fibroin (SF)- a protein that is commonly referred to as the structural component of Bombyx mori silks- has proved to be one of the most promising materials in contemporary bioelectronics.(1) It is popular due to an unusual set of characteristics: it is biocompatible,

mechanically adjustable, biodegradable, optically clear, and can even be processed in water.(2) With the continuous shift of bioelectronic devices via the silicon-based rigid component into the soft, bio-integrated structure, which is being performed by skin and tissue, these characteristics allow silk to have a pronounced advantage over standard synthetic surfaces.(3) Silk fibroin is manufactured at the molecular scale by the



deposition of closely-packed crystalline β -sheets into a weaker, amorphous framework.(4) The unusual structure enables the researcher to precisely control the stiffness, elasticity, water stability, and degradation rate of the structure through just a controlled processing cycle.(5) The ease of casting, printing, patterning or molding SF with no harsh solvents makes it readily assume various forms such as thin flexible films, fibers, hydrogels, microneedles and even whole-biodegradable electronic platforms.(6) Such formats may be combined with rigid and soft conductive materials and therefore silk presents a flexible transition between biological tissues and electronic devices.(7) In bioelectronics, SF plays a number of functions simultaneously: an elastic platform to wearable and epidermal sensor, an implantable or transient circuit dielectric layer, an encapsulant that is biocompatible which shields delicate components and a dynamic substance, particularly one combined with conductive polymers, nanoparticles, or nanomaterials of carbon.(8) Such versatility has seen the formation of devices based on silk being used in skin mounted sensor, dissolvable implants, neural interfaces, optical and biochemical sensors, and even edible or environmentally friendly electronics.(9) Seeing how well it can fit its own into tissues, support delicate biomolecules, and dissolve under command, it also can be highly valuable to next generation health monitoring and transient diagnostic systems.(10) Nonetheless, there is one obstacle, which is long-term stability. Biodegradability is perfectly suited to a temporary or resorbable implant, although it restricts the application of unmodified SF in chronic implants or continuous physiological implants, where mechanical and dielectric characteristics need to be maintained over months or years.(11) This has made recent studies consider what can be done utilizing various methods like augmenting crystallinity, crosslinking chemicals and most interestingly oriented crystallization that can elevate water resistance and structural strength as well as electrical performance to a considerable degree.(12) This review emphasizes the increasing use of silk fibroin in bioelectronics, existing issues, and mechanisms to overcome the current challenges with potential future solutions, including the way orientation crystallization can get the long-term survival of the bioelectronic technologies, which can be implanted and sustained in the future.(13)



Figure 1 In this figure, the article is talking about silk fibroin (SF) as a very versatile biomaterial that can be modified in terms of its properties by applying various techniques, such as sol-gel processing, gel casting, precision film forming, and controlled extraction. Its principal properties are biocompatibility, biodegradability, water solubility, transparency and the capacity to carry drugs and as a result, this property has enabled it to be used in numerous ways. Some of the large application areas feature in the external sections include implantable medical devices (drug delivery, transient electronics, scaffolds), wearable devices (electronic skin, smart insoles, tattoos), energy harvesters (flexible TEGs and wearable PEGs), multi-channel sensors (humidity, temperature, strain), and flexible electronic systems (memristors, LED wearables, air filtration). Generally, SF represents a universal system of highly bioelectronic and biomedical technologies.

2.0 Silk Fibroin as a Functional Biomaterial for Bioelectronic Interfaces

Silk fibroin (SF) has become a versatile choice of bioelectronic interface since it has a unique property of biocompatibility, mechanical control, optical transparency, and mild, aqueous processing.(14) The hierarchical nature of its structure means that its crystals formed out of β -sheet are encased by a softer amorphous network providing researchers with finer control over the properties of their aids to include stiffness, elasticity, water stability, and degradation.(15) It allows SF to be flexible to ultra-soft, tissue-like electronics, as well as more traditional rigorous devices. Silk as a substrate has an outstanding flexibility and conformability, together



with the ability to be compatible with conventional microfabrication techniques. These can be used to design ultrathin wearable biosensors, neural interfaces and entirely biodegradable circuits.(16) By being either, a dielectric or an encapsulant, low dielectric constant, high breakdown strength and thickness, and capability to form seamless, consistent films, together with its low dielectric constant, high breakdown stress, and implantable devices, will make silk a reliable performer in the organic transistor, capacitive sensor, and implantable systems.(17) It is also possible to make SF assume active functions in combination with conductive polymers, carbon nanomaterials, metal nanoparticles, or ionic additives. These composites have the ability to support conductive or ionically active components broadening the scope of biomedical applications of silk into biosensing and soft electrode application, and therapeutic interfaces.(18) Since silk inherently stabilizes fragile biomolecules such as enzymes, antibodies, drugs, etc. it also facilitates integrated biochemical bioproducts sensing and bioactive electronics.(19) The next significant benefit of silk is that it has an excellent biocompatibility. It incorporates well into tissues, suppresses swelling, and assists to achieve stable chronic contact. Its versatility in creating a wide variety of structures in films, fibers, hydrogels, microneedles, porous scaffolds allow the designer to create interfaces at all scales as well as application requirements.(20) One drawback is also that SF is biodegradable, which is an advantage with transient electronics but puts long-term implants constrained. The existing measures of enhancing crystallinity, chemical crosslinking, and most importantly oriented crystallization are trying to enhance the water resistance as well as structural permanence.(21) These progresses are making silk fibroin a better candidate as a next-generation bioelectronic technology that is tissue amenable.(22)

2.1 Table Functional Roles and Advantages of Silk Fibroin in Bioelectronic Interfaces

Functional Role	Key Properties	Bioelectronic Applications
Substrate	<ul style="list-style-type: none"> Flexible and conformable Tunable mechanical modulus Compatible with microfabrication (printing, lithography) Aqueous, solvent-free processing(23) 	<ul style="list-style-type: none"> Wearable/epidermal sensors Ultrathin flexible circuits Neural recording platforms Biodegradable/transient electronics(24)
Dielectric Layer	<ul style="list-style-type: none"> Low dielectric constant High breakdown strength Uniform thin-film formation Moisture-resistant when crystallized(25) 	<ul style="list-style-type: none"> Organic field-effect transistors (OFETs) Capacitive sensors Implantable circuits(26) Energy storage or passive components(27)
Encapsulant/Packaging	<ul style="list-style-type: none"> Biocompatible and non-inflammatory Tunable dissolution rates Protects electronics from(28) moisture/enzymes Can stabilize biological molecules(29) 	<ul style="list-style-type: none"> Implantable electronics Transient devices with controlled degradation Drug-loaded electronics and biosensors(30)
Active Functional Material	<ul style="list-style-type: none"> Forms conductive composites with CNTs, graphene, PEDOT: PSS, metal nanoparticles(31,32) Ion-conductive silk hydrogels Stabilizes enzymes, antibodies, peptides(33) 	<ul style="list-style-type: none"> Chemical/biochemical sensors Ionotropic components Soft electrodes Optical and electrochemical biosensing(34)
Biointegration & Tissue Interface	<ul style="list-style-type: none"> Excellent biocompatibility Low inflammation and immune response(35) Matches soft tissue mechanics Supports long-term tissue contact(36) 	<ul style="list-style-type: none"> Neural and cardiac interfaces(37) Skin-mounted electronics Bioactive therapeutic devices(38)
Structural Formats	<ul style="list-style-type: none"> Films, fibers, hydrogels, microneedles, scaffolds Patternable and moldable architectures(39) 	<ul style="list-style-type: none"> Multi-scale interface engineering Microstructured sensors Flexible, implantable device platforms(40)
Biointegration & Tissue Interface	<p>Challenge: Intrinsic biodegradability limits long-term stability(41)</p> <p>Solutions: Crystallinity enhancement, crosslinking, oriented crystallization(42)</p>	<ul style="list-style-type: none"> Chronic implants Durable biosensing platforms(43)

3.0 Limitations of Conventional Silk-Based Bioelectronic Devices

Minimal Kontrol over the Kinetic Degradation in vivo. Though the rate of degradation of silk can be modified with the processes, crystallinity tuning, and crosslinking, it is not yet easy to predict how this composite will degrade within the body.(44) Physiological conditions are diverse with respect to the enzyme activity, pH, fluid flow, inflammation and mechanical forces.(45) Such variations imply that a device that is created to fade away at the end of a certain time span might either disintegrate prematurely or stay longer than it is supposed to.(46) It makes it difficult to design time-sensitive transient devices and difficult to get a regulatory approval where reproducibility is mandatory.(47) Complicated Interaction with Biospecies Fluids and Proteins. Silk fibroin is permeable to these proteins, electrolytes, and biomolecules in the tissues that are surrounding it.(48) This makes it more biocompatible, but presents a number of issues. Protein adsorption has the capability to sensitive the surface charge and disrupt



electrophysiological signals and the biofouling effectively attenuates the sensor accuracy over time as well.(49) It can be degraded by the rate of enzymes and biomolecules and swelling because of the fluid uptake can lead to delamination in multilayer structures.(50) Of particular concern are neural probes and biochemical sensors which demand relatively low noisy interfaces.(51) Problems of Achieving Long-Term Electrical Reliability. The sensitivity of silk to moisture provides numerous possibilities of electrical failure. Metal traces can be corroded as water gains access to it, and sodium ion can have an upsetting effect on conductive polymers;(52) recurrent swelling-shrinkage can result in the separation of conductive layers. Cyclic strain may crack the conductor composite. All these combined causes the effects of drift, increased noise levels, and ultimate loss of signal, which restricts the usefulness of untouched silk in longitudinal implants or in persistent monitoring apparatus.(53) Processing Thermal and Chemical Constraints. Being a protein substance, silk is not able to stand high temperatures and rough solvents. Denaturation or yellowing can be obtained during heating and conventional processing chemicals in microelectronics can damage the matrix. Technologies that involve metal deposition, etching, and photolithography usually demand modified technologies or protective layers so as not to compromise the structure.(54) These factors complicate the ability to wire silk to high-performance semiconductor or nanoscale-built fabrication methods of high-performance electronics. Restricted Mechanical Strength as a Dynamic Physiological Environmental Constraint.(55) Silk is elastic, yet it does not adjust to the extreme motions of such tissues as the heart, gut, or skeletal muscles.(56) Stretching or bending many times can wear out films made of silk faster than the same elastomers (PDMS or Ecoflex). Cyclic deformation can also cause removal of b-sheet structures resulting in mechanical weakening with time. These are some of the constraints to the conventional SF in devices that would need to operate in the high-strain environments that are nonstop. Limitations in High-Density Microelectrode Integration.(57) The recently developed bioelectronic systems are becoming more and more dependent on high density microelectrode arrays having their geometries that are specific. The sensitivity of electrodes, patterns, and dielectric properties to the swelling, softness, and dimensional change of silk in wet conditions may produce swollen electrodes, distorted patterns and changed dielectric.(58) This instability may end in signal cross- talking or short-circuits. This complicates

realisation of the micron scale resolution needed in neural recording or mapping with high compared to more dimensionally stable synthetic polymers. Scalability and Standardization Shortcoming. Massive manufacture of electronics made of silk is not without challenges owing to the natural variation.(59) The source of cocoon, and methods of extracting the silk together with the storage medium have Hongkwe on the molecular weight, purity, and content in the b-sheet of silk.(60) The differences may lead to observable changes in electrical, mechanical and degradation behavior. There are no standard specifications that are biomedical-grade silk fibroin therefore, reproducibility and commercial scaling can be more challenging. Federal and Translational Barriers.(61) Although silk can be used up to now because it is essentially biocompatible, hybrids that involve metals, carbon nanomaterials, MXenes, or other conductive fillers bring new safety issues. Each component has to be tested in terms of stability over the long term, toxicity, inflammatory effect, and harmless degradation product. Multi-material systems are also in need of massive mechanical, chemical and electrical reliability testing.(62) The bioresorbable electronics do not have a proper route of regulation, and thus, their translation to the clinic is a slow and tedious process.(63)

3.1 Table Key Limitations of Conventional Silk-Based Bioelectronic Devices

Limitation	Description / Impact on Bioelectronic Performance
Unpredictable Degradation Kinetics In Vivo	Degradation varies with enzyme levels, pH, inflammation, and mechanical forces; can lead to premature device failure or prolonged persistence; complicates design of transient implants and regulatory evaluation.(64)
Interactions with Biological Fluids and Proteins	Protein adsorption alters surface charge and signal fidelity; biofouling reduces sensor accuracy; enzymatic activity accelerates degradation; swelling causes delamination in multilayer devices.(65)
Poor Long-Term Electrical Reliability	Moisture ingress causes corrosion of metals and degradation of conductive polymers; swelling-shrinkage cycles disrupt interfaces;



	conductive composites crack under strain; results in drift, noise, and signal loss.(66)
Thermal and Chemical Processing Restrictions	Sensitive to heat and harsh solvents; prone to denaturation and structural damage during fabrication;(67) requires modified lithography and metallization techniques; limits compatibility with CMOS and high-resolution microfabrication.(68)
Limited Mechanical Endurance Under Dynamic Loads	Fatigues more quickly than elastomers under repeated deformation; β -sheet disruption weakens structure over time; unsuitable for tissues with large or continuous movement (heart, gut, skeletal muscle).(69)
Challenges in High-Density Microelectrode Integration	Swelling and softness cause electrode displacement and pattern distortion; dielectric instability may lead to short-circuits; difficult to achieve sub-20- μ m precision for high-resolution neural and electrophysiology devices.(70)
Scalability and Batch Variability	Natural source variability affects molecular weight, purity, β -sheet content; impacts electrical and mechanical properties; lack of standardized silk fibroin specifications limits reproducibility and commercial scaling.(71)
Regulatory and Translational Barriers	Hybrid devices with metals or nanomaterials require extensive biocompatibility and toxicology testing; multi-material architectures need rigorous reliability studies; (72)absence of established pathways slows clinical translation.(73)

4.0 Importance of Long-Term Stability in Chronic Biopotential Recording

Chronic biopotential recording systems are one of the most critical in terms of their long-term stability. These

implants can be as long as months or even years in the body serving tissues that are experiencing constant motion, recovery and adaptation.(74) It may be long-term EEG or ECoG monitoring, intracortical neural recording, cardiac electrophysiology, or chronic EMG tracking, the device has to be reliable to provide valid signals without drift, degrading, or failure with time.(75) The fundamental principle of long-term functionality lies at the capability to maintain constant electrical contact between the electrode and an adjacent tissue.(76) The signals of biopotential, particularly very small ones such as neural spikes, require a low-impedance interface that is always maintained.(77) A small dislocation of the electrode, a minor alteration in material characteristics or even natural alteration of local tissues may cause noise, signal distortion or ultimately failure. Accumulating over a long period of implantation, these little disturbances lead to one of the most challenging tasks of designing a chronic device such as the establishment of a stable electrode-tissue interface. Biological reaction of the body complicates it further.(78) All implanted materials result in a certain level of reaction. Inflammation, protein adposition, glial scarring in the brain, or fibrotic encapsulation of a device in muscles or a heart can take a long time (weeks or months) to develop. Such changes may physically isolate electrodes with target cells, or make the interface more resistant, undermining or distorting signal recordings.(79) In order to reduce these problems, long-term implants should not just be biocompatible, but soft, flexible and able to retain similar characteristics on their surfaces throughout their life. Implanted materials are also under a lot of stress in the internal environment of the body.(80) The degradation of the material may be caused by physiological fluids, repetitive movement, changes in temperature, and biochemical activity.(81) Typical ways of failure are swelling due to hydration, conductive layer corrosion, delamination, and mechanical fatigue due to repeated movement. In moisture sensitive or biodegradable materials such as silk fibroin these problems can occur much faster, since natural degradation can cause alteration of dielectric properties or loss of mechanical integrity before the device has achieved its target lifespan. Dielectric stability is another critical factor with respect to long-term reliability.(82) Such differences in the values of dielectric constant, swelling in the presence of moisture or early breakdown of dielectric may both distort the signals and create leakage currents undermining accuracy and safety. Since large arrays of electrodes have a high density, geometric variation on a small scale can cause space resolution errors or result in



increased cross-talk, with severe impact on long-term mapping resolution. Long-term stability is necessary not only to ensure the accuracy of the data, but also to ensure the comfort and safety of patients.(83) Surgical devices that malfunction early may need revision surgery, disrupt a continuous treatment or follow up or result in misinterpretation of clinical results.(84) Long-term stability has now emerged as a major challenge in commercializing bioelectronic technologies to actual clinical application as regulatory requirements are starting to look more closely at chronic performance data.(85) In general, stability in the long-term is important as it minimizes biological and mechanical complications, optimizes patient risk, and reduces signal fidelity, which is necessary to achieve successful implantable bioelectronic systems.(86) Enhancing chronic survivability, by means of better materials, interfaces, and architecture--are other areas of interest in the design of next-generation bioelectronic technologies.(87)

5.0 Oriented Crystallization Silk Fibroin. A Novel Physical Modification Approach

Oriented crystallization is acquiring significant momentum as one of the most promising physical approaches to improving the functionality of silk fibroin (SF) in the contemporary bioelectronic sensors and devices.(88) Traditional silk processing generally generates films and fibers that have randomly distributed β -sheet domains that cannot easily be controlled, and thus mechanical behavior, dielectric stability and long-term structural integrity are hard to control. Conversely, in oriented crystallization such β -sheet structures are rearranged into directionally improved networks.(89) This orientation is possible via a range of physical stimuli such as mechanical stretching, shear forces, directional freezing or the use of magnetic and electric fields that direct SF molecules into more regular and highly organized crystal assemblies.(90) The end product has better mechanical strength, better dimensional stability and higher predictability which is very vital in long-term or implantable bioelectronic systems.(91) This molecular alignment is not only mechanically reinforced to advantage the process. β -sheet networks made of high orientation have an important effect on enhancing the dielectric stability of silk to lower the moisture absorption rate, lower swelling, and also avoid random alterations of electrical properties under physiological conditions.(92) Such enhancements contribute to the sustained operation of the device, noise reduction, signal drift reduction, and a reduced risk of delamination which

are beneficial in applications like neural interfaces, wearable biosensors, and bioelectronic patches which need to endure constant motion and have contact with fluids.(93) It is noteworthy that oriented crystallization does not require the use of chemical crosslinkers or likely harmful additives, which helps the crystallization method to retain the intrinsic biocompatibility and safety profile of silk.(94) Altogether, oriented crystallization is a clean, bio-safe, and very efficient modification technique that makes silk fibroin a good candidate of next-generation bioelectronic implants.(95) This strategy will give a powerful route to efficient, dependable and declined bioelectronic contact development since it permits minute control of mechanical soundness, degradation Kinetics and diffusion of dielectric properties.(96)

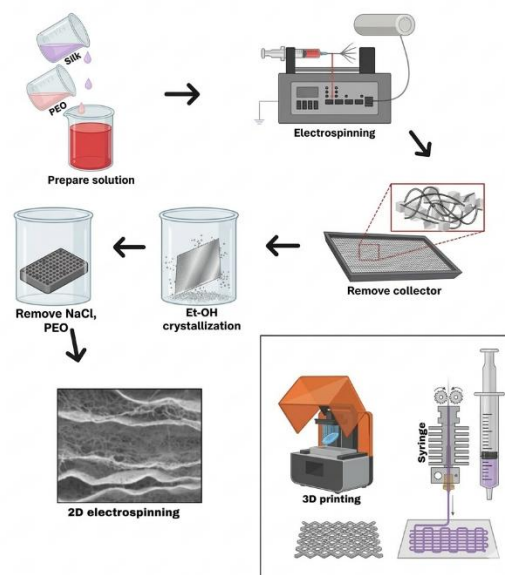


Figure 2

This image shows the sequential production of the nanofibrous materials made of silk fibroin (SF)-based materials through electrospinning. A homogeneous solution is first obtained by mixing silk fibroin with poly (ethylene oxide) (PEO). Electrospinning is then done on the solution to form fibrous mats deposited on a substrate. The fibers obtained are washed away using ethanol (EtOH) to cause crystallization and increase structural stability. PEO and salt (NaCl) are then removed to get pure porous silk fibroin structures. The material that is formed is in a nanofibrous form, as demonstrated in the SEM image. Also, the figure reveals the combination of electrospinning and 3D printing methods as a means of making structures more advanced.



6.0 Pre- Stretching Treatment as a Significant Method of Tunable Silk Membranes.

Before the treatment Pre-stretching is a powerful physical approach to the tuning of the structural and functional properties of tunable membranes of silk fibroin in bioelectronic applications.(97) Silk is made up of crystalline β -sheet domains and amorphous regions made up of random coils and α - helical structures giving it a hierarchical structure. When these molecular regions are in the relaxed state they are randomly oriented and hence offer medium mechanical flexibility but also make them more sensitive to moisture and mechanical deformation.(98) As a polymer film is pre-stretched, the chains of the polymer are oriented in the direction of imparted strain. This orientation facilitates the conversion of amorphous structure to β -sheet crystalline structures to a greater extent, as well as enhances intermolecular hydrogen bonding.(99) Consequently, the internal structure of the membrane is made more compact and anisotropic with great impact on its mechanical, electrical, and degradation properties.(100) Mechanically, pre-stretched silk membranes have been shown to have good tensile strength, Young's modulus and decreased creep deformation. The increased molecular packing increases the efficiency of load transfer in the material such that the membrane can endure the repetitive mechanical stress.(101) This is an important property of implantable bioelectronic devices that are in constant contact with physiological motions like cardiac pulsation or muscle contraction.(102) The structural organization is thus improved thus increasing durability and reliability of silk membranes in dynamic biological environments. The electrical behaviour of silk membranes is also pre-stretched. Higher crystallinity lowers the free volume of the polymer structure, and this restricts the uptake of water and thus a reduction in the variation of dielectric properties.(103) The β -sheet network with higher density enhances dielectric stability and lowers leakage currents which leads to similar electrical performance in the long term.(104) This stability is vital in the use of the material in neural interface, flexible biosensors and implantable electronic substrates where the material is required to transmit signals reliably.(105) One more significant benefit of pre-stretching is that it has an effect on degradation behavior. An increase in β -sheet content gives greater resistance to enzymatic and hydrolytic degradation, and extended life on the biological environment of silk membranes.(106) Through varying degrees of pre-stretching used in processing made material, researchers

can directly regulate the degradation rate of the material to meet the particular requirements of a biomedical application, such as temporary implants or long-term monitoring devices. Pre-stretching also enhances the dimensional stability of the silk membranes under aqueous situations.(107) Silk membranes are prone to untreated swelling when exposed to moisture, and it may lead to changes of the thickness, structural distortion, or delamination in multi-layer bioelectronic systems. Pre-stretched membranes in contrast are seen to swell less because of the tightly packed molecular structure.(108) This is used to maintain structural integrity and electrode and conductive layer alignment in high-resolution bioelectronic devices. Also, pre-stretching can improve interfacial adhesion between silk membranes and conductive materials including metal thin films and conductive polymers.(109) The enhanced surface stability lowers the tendency to form cracks and mechanical drift in case of repeated bending or cyclic loading thus improving the total life of the device.(110) As a result, in pre-stretching, a simple method is used to scale and achieve chemical-free and easy to tune performance characteristics in silk membranes. It can be optimized to have the maximum mechanical strength, flexibility, electrical stability, and degradation kinetics without compromising the intrinsic biocompatibility of silk fibroin through the control of strain levels and processing conditions.(111) Due to these reasons, pre-stretched silk membranes have great promises in the future of wearable and implantable bioelectronic systems.(112)

7.0 Silk Bioelectronics Crystallinity-Conductivity Relationship.

The long-term stability and electrical performance of bioelectronic systems based on silk fibroin highly depends on how the relationship between conductivity and crystallinity is. Silk fibroin is a semi-crystalline biopolymer which is made of highly ordered β -sheet crystalline structure embedded into disordered amorphous structure.(113) The relative area, tilt and the arrangement of these domains have a considerable influence on charge transportation, dielectric behavior, sensitivity to moisture, and interfacial electrical properties. Knowledge and management of this crystal-conductivity association is thus critical in boosting the optimal silk resources applicable as a substrate, dielectric lattice, encapsulation, and composite matrices in wearable and implantable bioelectronic devices.(114)

7.1 Crystallinity Structural Origin in Silk Fibroin



Silk Fibroin is a protein with a crystalline structure formed by the arrangement of protein molecules in a regular manner. <[human]>7.1 Structural Origin of Crystallinity in Silk Fibroin Silk Fibroin is a protein, whose crystalline structure is created by the arrangement of the protein molecules in a regular way.(115)Silk fibroin has a repeating amino acid sequence, which is the substance of the crystalline structure of this compound and is mainly made of glycine, alanine, and serine.(116) These amino acids help to form tightly packed structures of b-sheet that is stabilized by intermolecular hydrogen bonding. b-sheet domains resulting are nanocrystalline regions that allow rigidity of structure and high molecular packing density. The amorphous regions, on the contrary, are composed of random coil conformations and less organized helices that are more mobile and have increased free volume.(117) The general level of crystallinity of the fibroin is determined by different processing factors like the rate of drying, solvent treatment, mechanical stretching, thermal annealing, and exposure to humidity. With the growth of b-sheet formation, the speed of the structure decreases as well as the mobility of the molecules, resulting in the increase in crystallinity.(118)

7.2 Ionic and Insulating Conduction Electrical Conductivity of Silk.

Silk fibroin acts as a good electrical insulator in its dry form since it does not contain any free charge carriers required to conduct the electrons. But on exposure to moisture, especially at physiological conditions, ionic conduction may proceed by the absorption of water molecules and mobile ions.(119) The degree of crystallinity is very important in the regulation of this electrical behavior. The crystalline silk structures are highly crystallized which results in compact molecular packing to limit water absorption and decrease ionic mobility to minimize leakage currents and enhance electrical stability.(120) Less crystallinity, on the other hand, causes the free volume in the structure to be high, allowing more water to be absorbed and protons or ions transported. Consequently, silk materials have the property of insulating conduction as well as ionic conduction depending on their crystallinity and environmental humidity.(121)

7.3 The effect of crystallinity on the dielectric characteristics is discussed.

Dielectric behavior is an essential parameter of the bioelectronic system based on silk especially in capacitive sensors, neural interfaces and implantable

circuit.(122) The degree of crystallinity has a direct effect on several dielectric properties, and they are the dielectric constant, dielectric loss, breakdown voltage, and frequency stability.(123) The higher the crystallinity, the higher the restriction of dipolar movement of the polymer chains resulting in reduced dielectric loss and oscillations of polarization. Increased crystallinity also enhances dielectric strength, and also resistance to electrical breakdown.(124) This means that the higher the crystalline content of the silk membranes, the more stable the impedance profiles in the presence of aqueous conditions, and this is critical to the chronic implantable devices that need to be signal-reliable over the long term.(125)

7.4 Crystallinity in Conductive Composites made of Silk.

Silk fibroin has also been used in conjunction with conductive polymers, carbon nanomaterials, metallic nanoparticles and other conductive substances as electrically active composites in several bioelectronic devices.(126) The crystal arrangement of the silk kelp plays a major role on the method of transporting the charge in these composites. Crystallinity makes it stronger as far as mechanics are concerned, and eliminates the development of microcracks that may interfere with the conduction routes.(127) Also, the ordered b-sheet structures give better interfacial bonding between the silk matrix and conductive fillers, and thus effective transfer of charge. Excessive crystallinity can however limit polymer chain movement and influence the dispersion of fillers.(128) Thus, a balance between the crystalline and amorphous areas should be considered optimal to ensure the mechanical or mechanical stability is ensured as well as efficient electrical conductivity.(129)

7.5 Interaction of moisture and electrical drift.

The uptake of moisture is significant in dictating the electrical properties of the silk materials.(130) The amorphous structures of silk can easily take up water, and this may enhance ionic conductivity, dielectric behavior, create impedance hysteresis, and hasten the degradation of the material.(131) The attainment of greater crystallinity decreases the diffusion routes of water in the structure and, consequently, minimizes the swelling and stabilizes electrical parameters over time.(132) This moisture insensitivity is specifically significant in the case of implantable bioelectronics that can be used in physiological fluids over long durations.(133)



7.6 Charge Transport Mechanisms Psychological Processes.

The charge transport in silk based electronic systems may occur in a variety of ways, including ionic conduction by the presence of moisture, electronic conduction by conductive fillers and inter-facial polarization at material interfaces.(134) The high crystallinity usually inhibits the ionic conduction by reducing the intake of water and enhancing the preservative electronic conduction routes in the composites.(135) Thus, the best degree of crystallinity depends on the proposed device operation. Crystallinity Dielectric layers and insulating substrates should be high-crystallinity, ionic sensor use could be moderate-crystallinity, and balanced-crystallinity is necessary with flexible electrode systems that need mechanical and electrical stability.(136)

7.8 Trade-Offs and Strategies of Optimization.

Even though the increase in crystallinity enhances the mechanical strength and electrical stability, overcrystallization may decrease the flexibility and make the material brittle.(137) In order to obtain the best performance, researchers use many processing strategies to regulate crystallinity, such as the controlled mechanical stretching, gradual evaporation of solvent, mild thermal treatment and spatial crystallinity engineering.(138) The methods permit the accurate regulation of crystalline composition without reducing the viability of silk fibroin in biocompatibility.(139)

8.0 Chronic Biopotential Chronic Electronic Interfaces to a chronic chronicle.

Electronic interfaces made of oriented-crystalline (OC) silk represent a high-tech category of biointegrated materials that can be used to improve the mechanical and electrical stability of devices employed in chronic biopotential monitoring.(140) The processing of the silk fibroin membranes in these systems is performed to attain high level of molecular chain alignment and high 8-sheet crystallinity by controlled methods like mechanical stretching, directional drying and solvent or thermal annealing.(141) This structured crystalline structure is an enormous enhancement of mechanical strength, dimensional stability and moisture insensitivity of silk and therefore, is the most appropriate in long-term electrophysiological studies, which involve the use of electrical electrodes (electroencephalography (EEG), electrocardiography (ECG), electromyography (EMG)).(142) The level of intermolecular hydrogen

bonding, high-order 200 -sheet domains, and reduced free volume increase tensile strength and minimize swelling of OC silk membranes in physiological conditions.(143) Such structural properties reduce mechanical mismatch and delamination in the multilayer bioelectronically, and therefore they retain the structural integrity during repeated mechanical forces generated by the beating of the heart, muscle movement, or micromotion of the brain.(144) Besides mechanical advantages, OC silk has better electrical stability, hence, required to detect bioelectrical signals in the microvolt-millivolt.(145) Higher crystallinity leads to decreased ionic leakage, decreased dielectric loss, constant impedance over frequencies and higher dielectric breakdown strength.(146) The material prevents changes in ion-mediated conductivity and also limits water diffusion thereby maintaining uniform electrode-tissue impedance and enhances the signal to noise ratio during long-term recordings.(147) Mechanical compatibility with soft biological tissues is also another important benefit of OC silk interfaces. In contrast to hard electronic materials, OC silk provides an equal balance between strength and flexibility enabling the interface to be strongly adherent to tissue surfaces and to be resistant to fatigue caused by repeated deformation.(148) This flexibility minimizes micromotion at the electrode tissue interface and, therefore, decreases inflammatory reactions and fibrotic encapsulation which, otherwise, would deteriorate signal quality over time.(149,150) Moreover, OC silk membranes can be used as substrates or encapsulation surfaces of conductive materials that include thin metal electrode, conductive polymers and carbon-based nanomaterials.(151) The crystalline ordered network of the interfaces increases interfacial adhesion between the silk and conductive layers, minimizing cracks or delaminations during the long term functioning of the device and guarantees constant conduction of electrical signals in flexible detection devices.(152) Chronic bioelectronic devices are used in wet conditions with high concentrations of electrolytes, and thus, low water uptake and swelling of highly crystalline OC silk further enhance the stability in the long term, reducing the effects of electrochemical degradation and impedance shift.(153) Although being more crystalline, OC silk still has the same biocompatibility and biodegradability of silk fibroin, and the degradation rate can be accurately regulated by the content of β -sheet.(154) Increased crystallinity will reduce degradation over the long term, but moderate crystallinity will provide the possibility of controlled bioresorption into temporary implants, which do not



require surgical removal.(155) Because of this set of mechanical longevity, electrical stability, water resistance, and tunable biodegradability, OC silk-based electronic interfaces have great potential in chronic biopotential monitoring, such as long-term neural signal recordings, implantable cardiac rhythm monitoring, wearable EEG and EMG systems, and flexible epidermal electrodes, and allow the continuous and reliable recording of physiological signals in the long term.(156)

9.0 Biocompatibility and In Vivo Performance of Silk Interfaces that are modified.

A paramount aspect in assessment of the silk fibroin (SF)-based bioelectronic interfaces is biocompatibility and in vivo performance particularly in biomedical implantable apparatus and long-term physiological monitoring system.(157) Altered silk interfaces made by using methods like pre-stretching, an oriented crystallization process, solvent annealing and composite incorporation have been created not only to improve mechanical and electrical functionality, but to maintain the high level of biological compatibility that silk itself has. Silk fibroin has been perceived to possess low immunogenicity, low cytotoxicity, high hemocompatibility and biodegradability is regulateable making it one of the most appropriate biomedical integrations.(158) Being a protein-based biomaterial, which is primarily based on amino acids, including glycine, alanine, and serine, silk fibroin may be slowly broken down by natural body enzymes and broken down into non-toxic by-products.(159) Silk, in contrast to many synthetic polymers, does not release toxic acidic products of degradation which could inflame or irritate areas of implantation. Significantly, the structural changes enhancing the crystallinity of the β -sheets are mainly those that affect physical structure of the material as opposed to those that affect chemical composition, thus in this case the material can maintain its native biocompatibility despite the processing changes to enhance the device functionality.(160,161) The effect of biological response to modified silk interfaces is highly dependent on the physical properties of the membrane such as surface roughness, balance between hydrophilicity and hydrophobicity, crystallinity and mechanical stiffness.(162) These parameters control the cell adhesion, proliferation, and differentiation on the material surface. Moderately crystallized silk membranes have been found to offer the best conditions to promote cell attachment in that they are mechanically stable enough and at the same time retain adequate surface wettability to adsorb proteins which is crucial to

cellular interaction. Higher crystallinity does not tend to prevent cell adhesion needed proteins adsorption, but a slight decrease in water uptake is evident.(163) Various experimental works have shown that modified silk interfaces maintain high cell viability, generate low concentration of inflammatory cytokine production, and enhance positive tissue integration and less fibrotic capsule formation than many other traditional synthetic implant materials.(164,165) These biological reactions are especially beneficial in neural and cardiac bioelectronic devices that require the stable long-term tissue integration of the bioelectronics to be used in order to obtain reliable signal access. Another critical parameter of the performance of the silk-based bioelectronic systems is the in vivo degradation behavior.(166) The rate of degradation of silk fibroin highly relies on the crystalline structure and the proportion of 2-sheets.(167) Less crystallinity also means that the materials are more easily degraded as their amorphous regions are easier to attack by enzymes and water and more crystalline structures degrade slower and more predictably. In long-term bioelectronic use, degradation should be controlled since the material should have structural and functional integrity during the necessary period of use of the device.(168) Silk interfaces can thus be modified to stay stable over a period of weeks or months before breaking down to non-toxic compounds which can be recycled by the surrounding tissues.(169) This can be particularly useful in temporary biomedical implants and bioresorbable electronic systems, which are set to work over a limited time and then harmlessly vanish, without having to undergo surgical procedures.(170) Besides the biological compatibility and control of degradation, the electrical environment in physiological environments is also very important in the reliability of the implantable bioelectronics. The devices in vivo are constantly subjected to wet conditions, ionic body fluid, mechanical movement, and temperature change some of which can alter electrical characteristics.(171) Enhanced crystallinity and modified silk membranes show less swelling with moisture, constant impedance properties, low leakage currents and time-dependent stable signal-to-noise ratios.(172) These properties assist in stabilizing the electrical activity of long-term recording of biopotentials, e.g. neural activity or cardiogenic rhythms. The other critical consideration to mechanical devices that are implanted in dynamic tissues like the brain, heart or skeletal muscles where deformation of the devices takes place through physiological movement.(173) Silk interfaces that have been modified are characterized by



high-fatigue performance, great crack propagation resistance, constant adhesion performance with conductive layers, and low mechanical drift under extended conditions. Oriented-crystalline or pre-stretched silk structures in specific ensure structural reliability during cyclic loading conditions, enhancing the reliability of flexible implantable devices to a great extent.(174) Hemocompatibility and tissue compatibility are also noted to be relevant in applications that are in direct contact with blood or vascular tissues. Silk fibroin generally exhibits low platelet activation, low thrombus formation and good relations with endothelial cells, which implies a good blood compatibility.(175) Structural changes that enhance mechanical or electrical functionality do not add foreign chemical additives, and the interface made of silk can remain in contact with biological systems without changing its natural compatibility with the biological system and produces few chances of clotting or immune response.(176) Moreover, the data with modified silk interfaces can facilitate gradual tissue remodelling and alignment with adjacent biological structures, which make it possible to create stable bioelectronic interfaces during long implantation periods.(177) The possibility to blend the silk fibroin with bioactive molecules, growth factors, or antimicrobial agents is another benefit of this material as it can additionally improve the cellular responses and lower the risk of infection when the implantation occurs.(178) Furthermore, silk materials are optically transparent and permeable to some gases, which could be further used as the means of implementing sensing or diagnostic capabilities in multifunctional bioelectronic systems. Finally, the successful combination of the biological compatibility, mechanical stability, and electrical stability is the key to the long-term functional performance of implantable bioelectronic devices.(179) Engineered silk interfaces have constant electrode tissue contacts, low signal drift, controlled degradation, and low chance of device failure in long term implantation. Structural tunability combined with mechanical flexibility, electrical stability and natural biological safety makes modified silk fibroin a highly promising material platform in the next generation implantable and wearable bioelectronic technology, especially in chronic neural monitoring, cardiac sensing, advanced biosensors, and long term physiological recording systems.(180)

10. Applications and Future Perspectives

Bioelectronic interfaces based on modified silk fibroin do not only promise great opportunities in the contemporary biomedical devices but also introduce new

possibilities in the future developments of healthcare technologies. The fact that they can combine structural strength with softness enables them to mediate the mechanical discontinuity between hard electronic devices and fragile biological tissues.(181) The property is especially useful in implants in the long-term neural implants, cardiac patches and soft epidermal sensors where mechanical mismatch is likely to cause inflammation or signal instability.(182) Through the precise modulation of β -sheet crystallinity, the electrical insulation, moisture resistance as well as degradation can be manipulated and so custom silk membranes can be tailored to particular clinical duration and functional conditions. Other applications in addition to conventional biopotential monitoring, there are investigations into silk-based platforms to use as integrated sensing platforms that can monitor electrical, chemical and mechanical signals simultaneously.(183) As an example, multimodal biosensors can be built on silk substrates to measure electrophysiological activity and pH, temperature or metabolic changes. They are also optically transparent and therefore can be combined with optoelectronic systems including optical stimulation or imaging-assisted neural interfaces. Moreover, innovations in microfabrication and additive manufacturing could also facilitate microfabricated and patterned silk electronics with microscale resolution, as well as high-density electrode array patterned interfaces with better spatial resolution in brain-machine interfaces. Future plans are projected to focus on scalable fabrication techniques, mixed material systems in addition to long-term clinical validation.(184) The use of biodegradable conductors and environmental processing technology may result in complete sustainability of electronic systems with a low environmental impact. Moreover, it is possible to imbue silk with wireless communication devices and energy-gathering elements to create self-powered implantable products that will be able to monitor their health constantly without needing a battery change.(185) With the ever-evolving interdisciplinary research, silk fibroin can be vastly replaced by a core in the design of safe, durable and adaptive bioelectronics in personalized medicine as well as long term therapeutic monitoring. In addition to biomedical applications of silk-based bioelectronics, bioresorbable medical electronics, smart drug delivery, and regenerative medicine are also emerging applications of silk-based bioelectronics.(186) Due to the ability of silk fibroin to stabilize sensitive biomolecules, researchers are seeking systems in which therapeutic agents, enzymes or growth factors are incorporated into



silk-based electronic systems. These versatile systems might be able to measure physiological cues and provide a regulated response to therapy at the same time, developing closed-loop bioelectronic therapies.(187) As an example, personalized and responsive healthcare technologies could be achieved by having implantable devices that react to abnormal neural or cardiac activity and release drugs in response to the signal detected.(188)The other potential avenue is the combination of silk fibroin and bendable and stretchable electronics to come up with next-generation wearable health systems. Silk-based epidermal electronics can adhere to the skin surface and the epidermal device can be used to continuously measure physiological parameters, including heart rate, muscle activity, hydration, and body temperature.(189) The systems can be active in long-distance health monitoring, rehabilitation, and early diagnosis. Moreover, optical transparency of silk offers the ability to combine it with optical sensors, photonic devices, bioimaging systems that have the potential to provide both electrical and optical monitoring of biological tissues simultaneously.(190)Silk-based bioelectronics is also growing in power with the development of nanotechnology. Graphene, carbon nanotubes, MXenes, and metallic nanowires used as nanomaterials in silk matrices can be very useful in increasing electrical conductivity, sensing sensitivity, and mechanical performance. These nanocomposites can then be hybridized creating biosensors that are highly sensitive and detect biochemical markers, neurotransmitters or metabolic compounds in very low concentrations.(191) Additionally, nanostructured interfaces based on silk can enhance electrode tissue interface which is of particular significance in neural recording and brain machine interface technology.Silk fibroin has become a candidate material in the environmentally-friendly and biodegradable electronic systems in the transient electronics field.(192) The artificial devices made of silk substrates and biodegradable conductors and semiconductors can then be dissolved once their purpose has been served, instead of requiring surgery to take out the implant in medical devices or discarding electronic waste in environmental sensors. These temporary electronics can be used in post-surgical monitoring, temporary neural stimulation and implantable diagnostic systems with a life expectancy of a set time before it can safely degrade in the body.(193)In spite of these developments, however, there are a number of obstacles that need to be overcome before bioelectronics based on silk can be used extensively in clinical practice. Mass

production techniques should be established, so that the quality of materials, reproducibility, and low costs of production can be achieved. Processing protocols of silk fibroin should also be standardized to keep the batch-to-batch variation of mechanical, electrical and degradation properties to a minimum.(194) Along with that, there is a need to conduct an extensive long-term in vivo research to thoroughly comprehend the long-term biological reaction to devices made of silk and to develop a dependable regulatory system in order to obtain a medical authorization.(195)Future studies will also consider the enhancement of structural engineering engineering techniques including oriented crystallization, controlled crosslinking and hierarchical microstructuring as a means of further enhancing mechanical stability and electrical stability.(196) It can be also integrated with wireless communication systems, micro-energy harvesting technology, and soft robotics that could result in autonomy of implantable devices that could continuously monitor and provide therapeutic intervention without the use of external power sources.(197,198) Finally, the combination of biomaterials science, nanotechnology, and bioelectronics will see the use of silk fibroin as one of the platforms of medical devices of the next generation that are safe, adaptable, and able to interact with living tissues over the long-term.(199)

References

1. Wang Y, Blasioli DJ, Kim HJ, Kim HS, Kaplan DL. Cartilage tissue engineering with silk scaffolds and human articular chondrocytes. *Biomaterials*. 2006;27(25). doi:10.1016/j.biomaterials.2006.03.050
2. Rodriguez MJ, Dixon TA, Cohen E, Huang W, Omenetto FG, Kaplan DL. 3D freeform printing of silk fibroin. *Acta Biomater*. 2018;71. doi:10.1016/j.actbio.2018.02.035
3. Franck D, Gil ES, Adam RM, Kaplan DL, Chung YG, Estrada CR, et al. Evaluation of Silk Biomaterials in Combination with Extracellular Matrix Coatings for Bladder Tissue Engineering with Primary and Pluripotent Cells. *PLoS One*. 2013;8(2). doi:10.1371/journal.pone.0056237
4. Omenetto FG, Kaplan DL. New opportunities for an ancient material. *Science*. 2010. doi:10.1126/science.1188936
5. Wang W, Mei L, Wang F, Pei B, Li X. The potential matrix and reinforcement materials for the



- preparation of the scaffolds reinforced by fibers or tubes for tissue repair. In: *Tissue Repair: Reinforced Scaffolds*. 2017. doi:10.1007/978-981-10-3554-8_2
- Chen L, Cao C, Zhong W, Wang Z, Lv L. Design and Fabrication of Flexible Silk Fibroin/Lanthanide Ion Membranes with Multifunctional Properties of Fluorescence, Humidity Sensitivity, and Conductivity. *ACS Appl Mater Interfaces*. 2025;17(37). doi:10.1021/acsami.5c12288
 - Zou P, Li X, Zhao H, Qu S, Jiang Z, Lei X, et al. Recent advances of silk-fibroin-based hydrogel in the field of antibacterial application. *Polymer International*. 2025. doi:10.1002/pi.6699
 - Kharaziha M, Scheibel T, Salehi S, Kharaziha M, Scheibel T, Salehi S. Multifunctional naturally derived bioadhesives: From strategic molecular design toward advanced biomedical applications-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>). *Prog Polym Sci*. 2024;150.
 - Li C, Guo J, Liu X, Wang S, Zhang Z, Zhang Y, et al. Robust Wearable Sensors Based on Silk Fibroin Hydrogels Enforced by Spherical Polyelectrolyte Brushes with Metal Nanoparticles. *Langmuir*. 2025;41(44). doi:10.1021/acs.langmuir.5c04109
 - Li Z, Xu S, Xu Z, Shu S, Liu G, Zhou J, et al. Enhancing cellular behavior in repaired tissue via silk fibroin-integrated triboelectric nanogenerators. *Microsyst Nanoeng*. 2024;10(1). doi:10.1038/s41378-024-00694-5
 - Yang J, Luo J, Liu H, Shi L, Welch K, Wang Z, et al. Electrochemically Active, Compressible, and Conducting Silk Fibroin Hydrogels. *Ind Eng Chem Res*. 2020;59(19). doi:10.1021/acs.iecr.0c00407
 - Mi HY, Li H, Jing X, He P, Feng PY, Tao X, et al. Silk and Silk Composite Aerogel-Based Biocompatible Triboelectric Nanogenerators for Efficient Energy Harvesting. *Ind Eng Chem Res*. 2020;59(27). doi:10.1021/acs.iecr.0c01117
 - Hua J, Huang R, Yu M, You R, Wang L, Yan S, et al. High-performance silk fibroin/hyaluronic acid interpenetrating network hydrogel microneedles for diabetes management. *Int J Biol Macromol*. 2025;298. doi:10.1016/j.ijbiomac.2025.140357
 - Ge W, Gao Y, Zeng Y, Yu Y, Xie X, Liu L. Silk Fibroin Microneedles Loaded with Lipopolysaccharide-Pretreated Bone Marrow Mesenchymal Stem Cell-Derived Exosomes for Oral Ulcer Treatment. *ACS Appl Mater Interfaces*. 2024;16(29). doi:10.1021/acsami.4c04804
 - Abou Taleb MF, Alkahtani A, Mohamed SK, Buwalda SJ, Boere KWM, Dijkstra PJ, et al. A comprehensive approach to in vitro functional evaluation of Ag/alginate nanocomposite hydrogels. *Carbohydr Polym*. 2015;72(1).
 - Yang Z, Huang L, Song X, Wang M, Han X, Guan W, et al. Properties of 3D-printed continuous silk fiber-reinforced poly(caprolactone). *RSC Adv*. 2025;15(19). doi:10.1039/d5ra01302j
 - Sheng H, Ma Y, Zhang H, Yuan J, Li F, Li W, et al. Integration of Supercapacitors with Sensors and Energy-Harvesting Devices: A Review. *Advanced Materials Technologies*. 2024. doi:10.1002/admt.202301796
 - Zhang Q, Soham D, Liang Z, Wan J. Advances in wearable energy storage and harvesting systems. *Med-X*. 2025. doi:10.1007/s44258-024-00048-w
 - Gomez-Gijon S, Ortiz-Gómez I, Rivadeneyra A. Paper-Based Electronics: Toward Sustainable Electronics. *Advanced Sustainable Systems*. 2025. doi:10.1002/adsu.202400486
 - Adeel M, Lee HS, Asif K, Smith S, Kurt H, Rizzolio F, et al. Micro-Supercapacitors for Self-Powered Biosensors. *Small Science*. 2024. doi:10.1002/smssc.202400096
 - Phan TTV, Santhamoorthy M, Thirupathi K, Lin MC, Kim SC, Kumarasamy K. A review of conducting polymer hydrogels: Synthesis and characterization, and their sensors, and energy harvesting applications. *Microchemical Journal*. 2025. doi:10.1016/j.microc.2025.115137
 - Li B, Zhang S, Xu L, Su Q, Du B. Emerging Robust Polymer Materials for High-Performance Two-Terminal Resistive Switching Memory. *Polymers*. 2023. doi:10.3390/polym15224374
 - Ling S, Zhang C, Zhang C, Teng M, Ma C, Gao J, et al. Facile synthesis of MXene–Polyvinyl alcohol hybrid material for robust flexible memristor. *J Solid State Chem*. 2023;318. doi:10.1016/j.jssc.2022.123731



24. Hamdani D. Konsep Pemikiran Pendidikan Islam. *Front Neurosci.* 2021;14(1).
25. Ana S. Qris dan era baru transaksi pembayaran 4.0. *Front Neurosci.* 2021;14(1).
26. Putri IM. ASUHAN KEPERAWATAN PADA TN.SI DENGAN CHRONIC OBSTRUCTIVE PULMONARY DISEASE (COPD) DI RUANG RAWAT INAP A RSUD KANJURUAN KEPANJEN. Undergraduate thesis, Universitas Muhammadiyah Malang. 2023;14(1).
27. GIRSANG JCP. Gambaran Perilaku Pencarian Pengobatan Penderita Demam Berdarah Dengue di Kecamatan Medan Selayang. *Front Neurosci.* 2023;14(1).
28. Utami DP. Pengertian Penelitian Dekriptif Kualitatif. *Front Neurosci.* 2021;14(1).
29. Affani AR. Tingkat Stres Akademik Pada Mahasiswa Dalam Pembelajaran Daring Selama Pandemi Covid-19. Fakultas Psikologi Universitas Muhammadiyah Malang. 2021;14(1).
30. Asri MN. Pengaruh Edukasi Gizi Menggunakan Media Booklet Terhadap Pengetahuan Dan Sikap Ibu Hamil Tentang Asi Eksklusif Untuk Pencegahan Stunting. Diploma thesis, Universitas Andalas. 2022;14(1).
31. Kundu B, Rajkhowa R, Kundu SC, Wang X. Silk fibroin biomaterials for tissue regenerations. *Advanced Drug Delivery Reviews.* 2013. doi:10.1016/j.addr.2012.09.043
32. Qian XQ, Zhang M, Wang HY. Progress of silk fibroin biomaterial use in oral tissue regeneration engineering. *Critical Reviews in Biotechnology.* 2025. doi:10.1080/07388551.2025.2472621
33. Zou S, Yao X, Shao H, Reis RL, Kundu SC, Zhang Y. Nonmulberry silk fibroin-based biomaterials: Impact on cell behavior regulation and tissue regeneration. *Acta Biomaterialia.* 2022. doi:10.1016/j.actbio.2022.09.021
34. Rajkhowa R, Tsuzuki T, Kundu B, Kundu SC, Wang X. Recent Innovations in Silk Biomaterials Silk fibroin biomaterials for tissue regenerations ☆. Article in *Journal of Fiber Bioengineering and Informatics.* 2013.
35. De Giorgio G, Matera B, Vurro D, Manfredi E, Galstyan V, Tarabella G, et al. Silk Fibroin Materials: Biomedical Applications and Perspectives. *Bioengineering.* 2024. doi:10.3390/bioengineering11020167
36. Nasrine A, Narayana S, Gulzar Ahmed M, Sultana R, Noushida N, Raunak Salian T, et al. Neem (Azadirachta Indica) and silk fibroin associated hydrogel: Boon for wound healing treatment regimen. *Saudi Pharmaceutical Journal.* 2023;31(10). doi:10.1016/j.jsps.2023.101749
37. Ma L, Dong W, Lai E, Wang J. Silk fibroin-based scaffolds for tissue engineering. *Frontiers in Bioengineering and Biotechnology.* 2024. doi:10.3389/fbioe.2024.1381838
38. Zhu S, Zhang Q, Xu X, Liu Z, Cheng G, Long D, et al. Recent Advances in Silk Fibroin-Based Composites for Bone Repair Applications: A Review. *Polymers.* 2025. doi:10.3390/polym17060772
39. Li D, Liang R, Wang Y, Zhou Y, Cai W. Preparation of silk fibroin-derived hydrogels and applications in skin regeneration. *Health Science Reports.* 2024. doi:10.1002/hsr.2.2295
40. Rajkhowa R, Tsuzuki T, Kundu B, Kundu SC, Wang X. Recent Innovations in Silk Biomaterials Controlling the photocatalytic and optical properties of zinc oxide nanoparticles View project Silk fibroin biomaterials for tissue regenerations ☆. Article in *Journal of Fiber Bioengineering and Informatics.* 2013;(March).
41. Lehmann T, Vaughn AE, Seal S, Liechty KW, Zgheib C. Silk Fibroin-Based Therapeutics for Impaired Wound Healing. *Pharmaceutics.* 2022. doi:10.3390/pharmaceutics14030651
42. Mohammadzadehmoghadam S, LeGrand CF, Wong CW, Kinnear BF, Dong Y, Coombe DR. Fabrication and Evaluation of Electrospun Silk Fibroin/Halloysite Nanotube Biomaterials for Soft Tissue Regeneration. *Polymers (Basel).* 2022;14(15). doi:10.3390/polym14153004
43. Wray LS, Hu X, Gallego J, Georgakoudi I, Omenetto FG, Schmidt D, et al. Effect of processing on silk-based biomaterials: Reproducibility and biocompatibility. *J Biomed Mater Res B Appl Biomater.* 2011;99 B(1). doi:10.1002/jbm.b.31875



44. Paladini F, Pollini M. Novel Approaches and Biomaterials for Bone Tissue Engineering: A Focus on Silk Fibroin. *Materials*. 2022. doi:10.3390/ma15196952
45. Saleem M, Rasheed S, Yougen C. Silk fibroin/hydroxyapatite scaffold: a highly compatible material for bone regeneration. *Science and Technology of Advanced Materials*. 2020. doi:10.1080/14686996.2020.1748520
46. Lin D, Li M, Wang L, Cheng J, Yang Y, Wang H, et al. Multifunctional Hydrogel Based on Silk Fibroin Promotes Tissue Repair and Regeneration. *Advanced Functional Materials*. 2024. doi:10.1002/adfm.202405255
47. Das JM, Behere I, Upadhyay J, Borah R, Ingavle G. Advancements in Silkworm-Derived Silk Fibroin Biomaterials for Peripheral Nerve Regeneration. *OBM Neurobiology*. 2025. doi:10.21926/obm.neurobiol.2501265
48. Yao X, Zou S, Fan S, Niu Q, Zhang Y. Bioinspired silk fibroin materials: From silk building blocks extraction and reconstruction to advanced biomedical applications. *Materials Today Bio*. 2022. doi:10.1016/j.mtbio.2022.100381
49. Branković M, Zivic F, Grujovic N, Stojadinovic I, Milenkovic S, Kotorcevic N. Review of Spider Silk Applications in Biomedical and Tissue Engineering. *Biomimetics*. 2024. doi:10.3390/biomimetics9030169
50. Aldahish A, Shanmugasundaram N, Vasudevan R, Alqahtani T, Alqahtani S, Mohammad Asiri A, et al. Silk Fibroin Nanofibers: Advancements in Bioactive Dressings through Electrospinning Technology for Diabetic Wound Healing. *Pharmaceuticals*. 2024. doi:10.3390/ph17101305
51. Wang D, Liu H, Fan Y. Silk fibroin for vascular regeneration. *Microscopy Research and Technique*. 2017. doi:10.1002/jemt.22532
52. Elango J, Lijnev A, Zamora-Ledezma C, Alexis F, Wu W, Marín JMG, et al. The relationship of rheological properties and the performance of silk fibroin hydrogels in tissue engineering application. *Process Biochemistry*. 2023. doi:10.1016/j.procbio.2022.12.012
53. Lyu Y, Liu Y, He H, Wang H. Application of Silk-Fibroin-Based Hydrogels in Tissue Engineering. *Gels*. 2023. doi:10.3390/gels9050431
54. Cheng G, Davoudi Z, Xing X, Yu X, Cheng X, Li Z, et al. Advanced Silk Fibroin Biomaterials for Cartilage Regeneration. *ACS Biomaterials Science and Engineering*. 2018. doi:10.1021/acsbomaterials.8b00150
55. Li M, You J, Qin Q, Liu M, Yang Y, Jia K, et al. A Comprehensive Review on Silk Fibroin as a Persuasive Biomaterial for Bone Tissue Engineering. *International journal of molecular sciences*. 2023. doi:10.3390/ijms24032660
56. Zhang T, Zhang R, Zhang Y, Kannan PR, Li Y, Lv Y, et al. Silk-based biomaterials for tissue engineering. *Advances in Colloid and Interface Science*. 2025. doi:10.1016/j.cis.2025.103413
57. Sun Z, Huang R, Lyu H, Yu X, Wang W, Li J, et al. Silk Acid as an Implantable Biomaterial for Tissue Regeneration. *Adv Healthc Mater*. 2023;12(28). doi:10.1002/adhm.202301439
58. Yao D, Wang T, Zhang X, Wang Y. High Concentration Crystalline Silk Fibroin Solution for Silk-Based Materials. *Materials*. 2022;15(19). doi:10.3390/ma15196930
59. Zhou L, Chen D, Wu R, Li L, Shi T, Shangguang Z, et al. An injectable and photocurable methacrylate-silk fibroin/nano-hydroxyapatite hydrogel for bone regeneration through osteoimmunomodulation. *Int J Biol Macromol*. 2024;263. doi:10.1016/j.ijbiomac.2024.129925
60. Bai S, Han H, Huang X, Xu W, Kaplan DL, Zhu H, et al. Silk scaffolds with tunable mechanical capability for cell differentiation. *Acta Biomater*. 2015;20. doi:10.1016/j.actbio.2015.04.004
61. Nguyen TP, Nguyen QV, Nguyen VH, Le TH, Huynh VQN, Vo DVN, et al. Silk fibroin-based biomaterials for biomedical applications: A review. *Polymers*. 2019. doi:10.3390/polym11121933
62. Zhou J, Wu N, Zeng J, Liang Z, Qi Z, Jiang H, et al. Chondrogenic Differentiation of Adipose-Derived Stromal Cells Induced by Decellularized Cartilage Matrix/Silk Fibroin Secondary Crosslinking Hydrogel Scaffolds with a Three-Dimensional Microstructure. *Polymers (Basel)*. 2023;15(8). doi:10.3390/polym15081868



63. Zhu J, Du Y, Backman LJ, Chen J, Ouyang H, Zhang W. Cellular Interactions and Biological Effects of Silk Fibroin: Implications for Tissue Engineering and Regenerative Medicine. *Small*. 2025. doi:10.1002/sml.202409739
64. Li W, Zhao Y, Cheng Z, Niu F, Ding J, Bai Y, et al. Fine-tuning of porous microchannelled silk fibroin scaffolds for optimal tissue ingrowth. *Mater Des*. 2025;251. doi:10.1016/j.matdes.2025.113711
65. Ansari AI, Ahmad Sheikh N, Kumar N. Mechanical and in vitro study of 3D printed silk fibroin and bone-based composites biomaterials for bone implant application. *Proc Inst Mech Eng H*. 2024;238(7). doi:10.1177/09544119241259071
66. Norouzi F, Bagheri F, Hashemi-Najafabadi S. Alendronate releasing silk fibroin 3D bioprinted scaffolds for application in bone tissue engineering: Effects of alginate concentration on printability, mechanical properties and stability. *Results in Engineering*. 2024;22. doi:10.1016/j.rineng.2024.102186
67. Pollini M, Paladini F. Bioinspired materials for wound healing application: The potential of silk fibroin. *Materials*. 2020. doi:10.3390/ma13153361
68. Eftekhari BS, Ashtari B, Jahani M, Afjeh-Dana E, Janmey PA, Simorgh S, et al. Silk Fibroin-Based Matrices for the Guidance of Cell Interaction, Tissue Regeneration, and Crosstalk. *Macromolecular Bioscience*. 2025. doi:10.1002/mabi.202400629
69. Farokhi M, Mottaghitalab F, Fatahi Y, Saeb MR, Zarrintaj P, Kundu SC, et al. Silk fibroin scaffolds for common cartilage injuries: Possibilities for future clinical applications. *European Polymer Journal*. 2019. doi:10.1016/j.eurpolymj.2019.03.035
70. Chouhan D, Mandal BB. Silk biomaterials in wound healing and skin regeneration therapeutics: From bench to bedside. *Acta Biomaterialia*. 2020. doi:10.1016/j.actbio.2019.11.050
71. Rafiei S, Ghanbari-Abdolmaleki M, Zeinali R, Heidari-Keshel S, Rahimi A, Royanian F, et al. Silk fibroin/vitreous humor hydrogel scaffold modified by a carbodiimide crosslinker for wound healing. *Biopolymers*. 2024;115(6). doi:10.1002/bip.23612
72. de Lartigue C, Belda Marín C, Fitzpatrick V, Esposito A, Casale S, Landoulsi J, et al. Silk Foams with Metallic Nanoparticles as Scaffolds for Soft Tissue Regeneration. *Int J Mol Sci*. 2024;25(22). doi:10.3390/ijms252212377
73. Bi X, Mao Z, Li L, Zhang Y, Yang L, Hou S, et al. Janus decellularized membrane with anisotropic cell guidance and anti-adhesion silk-based coatings for spinal dural repair. *Nature Communications*. 2025;16(1). doi:10.1038/s41467-025-56872-0
74. Lawrence BD. Methods to Produce Silk Fibroin Film Biomaterials for Applications in Corneal Tissue Regeneration. Ms Thesis. 2008.
75. Su X, Wei L, Xu Z, Qin L, Yang J, Zou Y, et al. Evaluation and Application of Silk Fibroin Based Biomaterials to Promote Cartilage Regeneration in Osteoarthritis Therapy. *Biomedicines*. 2023. doi:10.3390/biomedicines11082244
76. Kamaraj M, Rezayof O, Barer A, Kim H, Moghimi N, Joshi A, et al. Development of silk microfiber-reinforced bioink for muscle tissue engineering and in situ printing by a handheld 3D printer. *Biomaterials Advances*. 2025;166. doi:10.1016/j.bioadv.2024.214057
77. Jiang W, Xiang X, Song M, Shen J, Shi Z, Huang W, et al. An all-silk-derived bilayer hydrogel for osteochondral tissue engineering. *Mater Today Bio*. 2022;17. doi:10.1016/j.mtbio.2022.100485
78. Dutta R, Chowdhury S, Kar K, Mazumder K. Silk Fibroin-Based Biomaterial Scaffold in Tissue Engineering: Present Persuasive Perspective. *Regenerative Engineering and Translational Medicine*. 2025. doi:10.1007/s40883-024-00374-w
79. Yue M, Zhou Y, Li Z. Macrophage lysate-derived cytokine network combined with silk fibroin hydrogel promotes diabetic vascularized bone regeneration. *Chemical Engineering Journal*. 2024;490. doi:10.1016/j.cej.2024.151892
80. Veeman D, Sai MS, Sureshkumar P, Jagadeesha T, Natrayan L, Ravichandran M, et al. Additive Manufacturing of Biopolymers for Tissue Engineering and Regenerative Medicine: An Overview, Potential Applications, Advancements, and Trends. *International Journal of Polymer Science*. 2021. doi:10.1155/2021/4907027



81. Zhang F, Yang R, Zhang P, Qin J, Fan Z, Zuo B. Water-Rinsed Nonmulberry Silk Film for Potential Tissue Engineering Applications. *ACS Omega*. 2019;4(2). doi:10.1021/acsomega.8b03542
82. Rahman M, Dip TM, Nur MG, Padhye R, Houshyar S. Fabrication of Silk Fibroin-Derived Fibrous Scaffold for Biomedical Frontiers. *Macromolecular Materials and Engineering*. 2024. doi:10.1002/mame.202300422
83. Yodmuang S, McNamara SL, Nover AB, Mandal BB, Agarwal M, Kelly TAN, et al. Silk microfibril-reinforced silk hydrogel composites for functional cartilage tissue repair. *Acta Biomater*. 2015;11(1). doi:10.1016/j.actbio.2014.09.032
84. Strenge JT, Smeets R, Nemati F, Fuest S, Rhode SC, Stuermer EK. Biodegradable Silk Fibroin Matrices for Wound Closure in a Human 3D Ex Vivo Approach. *Materials*. 2024;17(12). doi:10.3390/ma17123004
85. Jyoti T N, Rudrakshi C, M L V P. A New Horizon in Biocompatible Membrane: The Role of Collagen and Silk Fibroin in Tissue Regeneration-Case series. *Journal of Biology and Medicine*. 2024;8(1). doi:10.17352/jbm.000043
86. Ma H, Xie B, Chen H, Song P, Zhou Y, Jia H, et al. High-strength and high-elasticity silk fibroin-composite gelatin biomaterial hydrogels for rabbit knee cartilage regeneration. *Front Mater*. 2024;11. doi:10.3389/fmats.2024.1390372
87. Liu J, Huang R, Li G, Kaplan DL, Zheng Z, Wang X. Generation of Nano-pores in Silk Fibroin Films Using Silk Nanoparticles for Full-Thickness Wound Healing. *Biomacromolecules*. 2021;22(2). doi:10.1021/acs.biomac.0c01411
88. Shi W, Sun M, Hu X, Ren B, Cheng J, Li C, et al. Structurally and Functionally Optimized Silk-Fibroin-Gelatin Scaffold Using 3D Printing to Repair Cartilage Injury In Vitro and In Vivo. *Advanced Materials*. 2017;29(29). doi:10.1002/adma.201701089
89. Sabarees G, Tamilarasi GP, Velmurugan V, Alagarsamy V, Sibuh BZ, Sikarwar M, et al. Emerging trends in silk fibroin based nanofibers for impaired wound healing. *Journal of Drug Delivery Science and Technology*. 2023. doi:10.1016/j.jddst.2022.103994
90. Gundogdu G, Okhunov Z, Starek S, Veneri F, Orabi H, Holzman SA, et al. Evaluation of Bi-Layer Silk Fibroin Grafts for Penile Tunica Albuginea Repair in a Rabbit Corporoplasty Model. *Front Bioeng Biotechnol*. 2021;9. doi:10.3389/fbioe.2021.791119
91. Bal-Öztürk A, Alarçin E, Yaşayan G, Avci-Adali M, Khosravi A, Zarepour A, et al. Innovative approaches in skin therapy: bionanocomposites for skin tissue repair and regeneration. *Materials Advances*. 2024. doi:10.1039/d4ma00384e
92. Fazal N, Latief N. Bombyx mori derived scaffolds and their use in cartilage regeneration: a systematic review. *Osteoarthritis and Cartilage*. 2018. doi:10.1016/j.joca.2018.07.009
93. Khosropanah MH, Rajabi M, Behboodi Tanourlouee S, Azimzadeh A, Alizadeh Vaghasloo M, Hassannejad Z. Silk as a promising biomaterial for 3D bioprinting: a comprehensive review of Bombyx mori silk's biomedical applications. *International Journal of Polymeric Materials and Polymeric Biomaterials*. 2025. doi:10.1080/00914037.2024.2344608
94. Ansari AI, Sheikh NA, Kumar N, Nath J. Three-dimensional printed silk fibroin and fenugreek based bio-composites scaffolds. *Proceedings of the Institution of Mechanical Engineers, Part L: Journal of Materials: Design and Applications*. 2024;238(11). doi:10.1177/14644207241241156
95. Grabska-Zielińska S, Sionkowska A, Carvalho Â, Monteiro FJ. Biomaterials with potential use in bone tissue regeneration-collagen/chitosan/silk fibroin scaffolds cross-linked by EDC/NHS. *Materials*. 2021;14(5). doi:10.3390/ma14051105
96. Das JM, Upadhyay J, Monaghan MG, Borah R. Impact of the Reduction Time-Dependent Electrical Conductivity of Graphene Nanoplatelet-Coated Aligned Bombyx mori Silk Scaffolds on Electrically Stimulated Axonal Growth. *ACS Appl Bio Mater*. 2024;7(4). doi:10.1021/acsabm.4c00052
97. Gao Y, Wang Y, Zhang J, Zhang M, Dai C, Zhang Y, et al. Advancing neural regeneration via adaptable hydrogels: Enriched with Mg²⁺ and silk fibroin to facilitate endogenous cell infiltration and macrophage polarization. *Bioact Mater*. 2024;33. doi:10.1016/j.bioactmat.2023.10.026



98. Lee WJ, Cho K, Jung G, Kim AY, Kim GW. The osteogenic effects of sponges synthesized with biomaterials and nano-hydroxyapatite. *Biomed Phys Eng Express*. 2023;9(4). doi:10.1088/2057-1976/acdb7d
99. Bharadwaz A, Jayasuriya AC. Recent trends in the application of widely used natural and synthetic polymer nanocomposites in bone tissue regeneration. *Materials Science and Engineering C*. 2020. doi:10.1016/j.msec.2020.110698
100. Ghosh S, Pati F. Decellularized extracellular matrix and silk fibroin-based hybrid biomaterials: A comprehensive review on fabrication techniques and tissue-specific applications. *International Journal of Biological Macromolecules*. 2023. doi:10.1016/j.ijbiomac.2023.127410
101. Woloszyk A, Buschmann J, Waschkies C, Stadlinger B, Mitsiadis TA. Human dental pulp stem cells and gingival fibroblasts seeded into silk fibroin scaffolds have the same ability in attracting vessels. *Front Physiol*. 2016;7(APR). doi:10.3389/fphys.2016.00140
102. Zhao YH, Niu CM, Shi JQ, Wang YY, Yang YM, Wang HB. Novel conductive polypyrrole/silk fibroin scaffold for neural tissue repair. *Neural Regen Res*. 2018;13(8). doi:10.4103/1673-5374.235303
103. Deng X, Gould M, Ali MA. A review of current advancements for wound healing: Biomaterial applications and medical devices. *Journal of Biomedical Materials Research - Part B Applied Biomaterials*. 2022. doi:10.1002/jbm.b.35086
104. Yao D, Liu H, Fan Y. Silk scaffolds for musculoskeletal tissue engineering. *Experimental Biology and Medicine*. 2016. doi:10.1177/1535370215606994
105. Radulescu DM, Andronescu E, Vasile OR, Ficai A, Vasile BS. Silk fibroin-based scaffolds for wound healing applications with metal oxide nanoparticles. *Journal of Drug Delivery Science and Technology*. 2024. doi:10.1016/j.jddst.2024.105689
106. Huang X, Zheng Y, Ming J, Geng A, Liu L, Zheng J, et al. Bioinspired poly(lactic acid)/silk fibroin-based dressings with wireless electrical stimulation and instant self-adhesion for promoting wound healing. *Ind Crops Prod*. 2025;223. doi:10.1016/j.indcrop.2024.120179
107. Zhu W, Wang H, Feng B, Liu G, Bian Y, Zhao T, et al. Self-Healing Hyaluronic Acid-based Hydrogel with miRNA140-5p Loaded MON-PEI Nanoparticles for Chondrocyte Regeneration: Schiff Base Self-Assembly Approach. *Advanced Science*. 2025;12(1). doi:10.1002/advs.202406479
108. Murugapandian R, Mohan SG, Sridhar TM, Nambi Raj NA, Uthirapathy V. Comparative Analysis of Electrospun Silk Fibroin/Chitosan Sandwich-Structured Scaffolds for Osteo Regeneration: Evaluating Mechanical Properties, Biological Performance, and Drug Release. *ACS Omega*. 2024;9(26). doi:10.1021/acsomega.4c01069
109. Zhang Y, Li M, Chang J, Li C, Hui Y, Wang Y, et al. Silk fibroin-gelatine haemostatic sponge loaded with thrombin for wound haemostasis and tissue regeneration. *Burns Trauma*. 2024;12. doi:10.1093/burnst/tkae026
110. Nosrati H, Pourmotabed S, Sharifi E. A review on some natural biopolymers and their applications in angiogenesis and tissue engineering. *Journal of Applied Biotechnology Reports*. 2018;5(3). doi:10.29252/JABR.05.03.01
111. Attri K, G. H. S, Gulabrao DP, Teja KSS, Garai I, Pandey AK, et al. Silk Biomaterials: Applications and Future Prospects in Biomedical Engineering. *UTTAR PRADESH JOURNAL OF ZOOLOGY*. 2024;45(16). doi:10.56557/upjoz/2024/v45i164301
112. Gong D, Zhai Q, Wu M, Tao X, Wang F. Promotion of rat femoral distal bone defect repair using alginate-silk fibroin composite hydrogel. *Int Immunopharmacol*. 2025;161. doi:10.1016/j.intimp.2025.114973
113. dos Santos FV, Siqueira RL, de Moraes Ramos L, Yoshioka SA, Branciforti MC, Correa DS. Silk fibroin-derived electrospun materials for biomedical applications: A review. *International Journal of Biological Macromolecules*. 2024. doi:10.1016/j.ijbiomac.2023.127641
114. Chung YG, Tu D, Franck D, Gil ES, Algarrahi K, Adam RM, et al. Acellular bi-layer silk fibroin scaffolds support tissue regeneration in a rabbit model of onlay urethroplasty. *PLoS One*. 2014;9(3). doi:10.1371/journal.pone.0091592



115. Bai S, Zhang M, Huang X, Zhang X, Lu C, Song J, et al. A bioinspired mineral-organic composite hydrogel as a self-healable and mechanically robust bone graft for promoting bone regeneration. *Chemical Engineering Journal*. 2021;413. doi:10.1016/j.cej.2020.127512
116. Agostinacchio F, Mu X, Dirè S, Motta A, Kaplan DL. In Situ 3D Printing: Opportunities with Silk Inks. *Trends in Biotechnology*. 2021. doi:10.1016/j.tibtech.2020.11.003
117. Khademolqorani S, Tavanai H, Chronakis IS, Boisen A, Ajallouei F. The determinant role of fabrication technique in final characteristics of scaffolds for tissue engineering applications: A focus on silk fibroin-based scaffolds. *Materials Science and Engineering C*. 2021;122. doi:10.1016/j.msec.2021.111867
118. Singh YP, Bandyopadhyay A, Dey S, Bhardwaj N, Mandal BB. Trends and advances in silk based 3D printing/bioprinting towards cartilage tissue engineering and regeneration. *Progress in Biomedical Engineering*. 2024. doi:10.1088/2516-1091/ad2d59
119. Ansari AI, Sheikh NA. Mechanical and In Vitro Analysis of 3D Printed Silk Fibroin/Bone/Polycaprolactone/Chitosan Composite Scaffolds. *Journal of The Institution of Engineers (India): Series C*. 2024;105(6). doi:10.1007/s40032-024-01096-2
120. Grabska-Zielińska S, Sionkowska A, Coelho CC, Monteiro FJ. Silk fibroin/collagen/chitosan scaffolds cross-linked by a glyoxal solution as biomaterials toward bone tissue regeneration. *Materials*. 2020;13(15). doi:10.3390/ma13153433
121. Escobar A, Serafin A, Carvalho MR, Culebras M, Cantarero A, Beaucamp A, et al. Electroconductive poly(3,4-ethylenedioxythiophene) (PEDOT) nanoparticle-loaded silk fibroin biocomposite conduits for peripheral nerve regeneration. *Adv Compos Hybrid Mater*. 2023;6(3). doi:10.1007/s42114-023-00689-2
122. Roy T, Maity PP, Rameshbabu AP, Das B, John A, Dutta A, et al. Core-shell nanofibrous scaffold based on polycaprolactone-silk fibroin emulsion electrospinning for tissue engineering applications. *Bioengineering*. 2018;5(3). doi:10.3390/bioengineering5030068
123. Watchararot T, Prasongchean W, Thongnuek P. Angiogenic property of silk fibroin scaffolds with adipose - Derived stem cells on chick chorioallantoic membrane. *R Soc Open Sci*. 2021;8(3). doi:10.1098/rsos.201618
124. Patel KD, Lamarra KA, Sawadkar P, Ludwig A, Perriman AW. Silk Fibroin/GelMA-Based Hydrogels as a Platform for Tissue Adhesives and Tissue Engineering. *ACS Biomaterials Science and Engineering*. 2025. doi:10.1021/acsbiomaterials.5c00286
125. Kundu B, Brancato V, Oliveira JM, Correlo VM, Reis RL, Kundu SC. Silk fibroin promotes mineralization of gellan gum hydrogels. *Int J Biol Macromol*. 2020;153. doi:10.1016/j.ijbiomac.2019.10.269
126. Yang J, Kim S, Hwang K, Lee J. Preparation and Characterization of Cellulose-Chitosan-Silk Fibroin Composite Films Using Lithium Bromide (LiBr) Solution. *Journal of the Korean Wood Science and Technology*. 2025;53(2). doi:10.5658/WOOD.2025.53.2.193
127. Alkazemi H, Chai J, Allardyce BJ, Lokmic-Tomkins Z, O'Connor AJ, Heath DE. Glycerol-plasticized silk fibroin vascular grafts mimic key mechanical properties of native blood vessels. *J Biomed Mater Res A*. 2025;113(1). doi:10.1002/jbm.a.37802
128. Bojedla SSR, Kattimani V, Alwala AM, Nikzad M, Masood SH, Riza S, et al. Augmented Repair and Regeneration of Critical Size Rabbit Calvaria Defects with 3D Printed Silk Fibroin Microfibers Reinforced PCL Composite Scaffolds. *Biomedical Materials and Devices*. 2023;1(2). doi:10.1007/s44174-023-00072-1
129. Qu J, Zhang H. Roles of Mesenchymal Stem Cells in Spinal Cord Injury. *Stem Cells International*. 2017. doi:10.1155/2017/5251313
130. Fan Z, Xiao L, Lu G, Ding Z, Lu Q. Water-insoluble amorphous silk fibroin scaffolds from aqueous solutions. *J Biomed Mater Res B Appl Biomater*. 2020;108(3). doi:10.1002/jbm.b.34434
131. Zhang Y, Sheng R, Chen J, Wang H, Zhu Y, Cao Z, et al. Silk Fibroin and Sericin Differentially Potentiate the Paracrine and Regenerative Functions of Stem Cells Through Multiomics



- Analysis. *Advanced Materials*. 2023;35(20). doi:10.1002/adma.202210517
132. Li J, Liu X, Tao W, Li Y, Du Y, Zhang S. Micropatterned compositemembrane guides oriented cell growth and vascularization for accelerating wound healing. *Regen Biomater*. 2023;10. doi:10.1093/rb/rbac108
133. Quan S, Yang J, Huang S, Shao J, Liu Y, Yang H. Silk fibroin as a potential candidate for bone tissue engineering applications. *Biomaterials Science*. 2024. doi:10.1039/d4bm00950a
134. Ansari AI, Sheikh NA. Biocompatible Scaffold Based on Silk Fibroin for Tissue Engineering Applications. *Journal of The Institution of Engineers (India): Series C*. 2023. doi:10.1007/s40032-022-00891-z
135. Zhang Q, Liu Z, He YY, Huang T, Yang X, Duan L, et al. Osteoimmunity-Regulating biospun 3D silk scaffold for bone regeneration in critical-size defects. *J Adv Res*. 2025. doi:10.1016/j.jare.2025.04.032
136. Subia B, Rao RR, Kundu SC. Silk 3D matrices incorporating human neural progenitor cells for neural tissue engineering applications. *Polym J*. 2015;47(12). doi:10.1038/pj.2015.69
137. Su P, Qian Y, You X, Zhang F, Shen Y. Aligned silk fibroin fiber conduits with enhanced capability for guiding peripheral nerve repair. *Eur J Med Res*. 2025;30(1). doi:10.1186/s40001-025-03030-3
138. Zhang HJ, Li FS, Wang F, Wang H, He TC, Reid RR, et al. Transgenic PDGF-BB sericin hydrogel potentiates bone regeneration of BMP9-stimulated mesenchymal stem cells through a crosstalk of the Smad-STAT pathways. *Regen Biomater*. 2023;10. doi:10.1093/rb/rbac095
139. Hong H, Lee OJ, Lee YJ, Lee JS, Ajiteru O, Lee H, et al. Cytocompatibility of modified silk fibroin with glycidyl methacrylate for tissue engineering and biomedical applications. *Biomolecules*. 2021;11(1). doi:10.3390/biom11010035
140. Wibowo UA, Judawisastra H, Barlian A, Alfarafisa NM, Moegni KF, Remelia M. Development of salt leached silk fibroin scaffold using direct dissolution techniques for cartilage tissue engineering. *Int J Adv Sci Eng Inf Technol*. 2019;9(3). doi:10.18517/ijaseit.9.3.4511
141. You H, Zhang Q, Yan S, You R. All-aqueous-processed Silk Fibroin/Chondroitin Sulfate Scaffolds. *Fibers and Polymers*. 2021;22(11). doi:10.1007/s12221-021-1422-y
142. DeBari MK, Niu X, Scott J V., Griffin MD, Pereira SR, Cook KE, et al. Therapeutic Ultrasound Triggered Silk Fibroin Scaffold Degradation. *Adv Healthc Mater*. 2021;10(10). doi:10.1002/adhm.202100048
143. Wang L, Zhang Y, Xia Y, Xu C, Meng K, Lian J, et al. Photocross-linked silk fibroin/hyaluronic acid hydrogel loaded with hDPSC for pulp regeneration. *Int J Biol Macromol*. 2022;215. doi:10.1016/j.ijbiomac.2022.06.087
144. Liu W, Zhou Z, Zhang S, Shi Z, Tabarini J, Lee W, et al. Precise Protein Photolithography (P3): High Performance Biopatterning Using Silk Fibroin Light Chain as the Resist. *Advanced Science*. 2017;4(9). doi:10.1002/advs.201700191
145. Naskar D, Bhattacharjee P, Ghosh AK, Mandal M, Kundu SC. Carbon Nanofiber Reinforced Nonmulberry Silk Protein Fibroin Nanobiocomposite for Tissue Engineering Applications. *ACS Appl Mater Interfaces*. 2017;9(23). doi:10.1021/acsami.6b04777
146. Zheng D, Chen T, Han L, Lv S, Yin J, Yang K, et al. Synergetic integrations of bone marrow stem cells and transforming growth factor- β 1 loaded chitosan nanoparticles blended silk fibroin injectable hydrogel to enhance repair and regeneration potential in articular cartilage tissue. *Int Wound J*. 2022;19(5). doi:10.1111/iwj.13699
147. Weitkamp JT, Wöltje M, Nußpickel B, Schmidt FN, Aibibu D, Bayer A, et al. Silk fiber-reinforced hyaluronic acid-based hydrogel for cartilage tissue engineering. *Int J Mol Sci*. 2021;22(7). doi:10.3390/ijms22073635
148. Zhao Z hu, Ma X long, Ma J xiong, Kang J yu, Zhang Y, Guo Y. Sustained release of naringin from silk-fibroin-nanohydroxyapatite scaffold for the enhancement of bone regeneration. *Mater Today Bio*. 2022;13. doi:10.1016/j.mtbio.2022.100206
149. Jao D, Mou X, Hu X. Tissue Regeneration: A Silk Road. *J Funct Biomater*. 2016;7(3). doi:10.3390/jfb7030022



150. Umuhoza D, Yang F, Long D, Hao Z, Dai J, Zhao A. Strategies for Tuning the Biodegradation of Silk Fibroin-Based Materials for Tissue Engineering Applications. *ACS Biomaterials Science and Engineering*. 2020. doi:10.1021/acsbomaterials.9b01781
151. Costa JB, Silva-Correia J, Oliveira JM, Reis RL. Fast Setting Silk Fibroin Bioink for Bioprinting of Patient-Specific Memory-Shape Implants. *Adv Healthc Mater*. 2017;6(22). doi:10.1002/adhm.201701021
152. Kheiri L, Golestaneh A, Mehdikhani M, Razavi SM, Etemadi N. Histological Evaluation of Subcutaneous Tissue Reactions to a Novel Bilayer Polycaprolactone/Silk Fibroin/Strontium Carbonate Nanofibrous Membrane for Guided Bone Regeneration: A Study in Rabbits. *Clin Exp Dent Res*. 2025;11(3). doi:10.1002/cre2.70140
153. Lee MC, Kim DK, Lee OJ, Kim JH, Ju HW, Lee JM, et al. Fabrication of silk fibroin film using centrifugal casting technique for corneal tissue engineering. *J Biomed Mater Res B Appl Biomater*. 2016;104(3). doi:10.1002/jbm.b.33402
154. Wong K, Tan XH, Li J, Hui JHP, Goh JCH. An In Vitro Macrophage Response Study of Silk Fibroin and Silk Fibroin/Nano-Hydroxyapatite Scaffolds for Tissue Regeneration Application. *ACS Biomater Sci Eng*. 2024;10(11). doi:10.1021/acsbomaterials.4c00976
155. Sahu N, Baligar P, Midha S, Kundu B, Bhattacharjee M, Mukherjee S, et al. Nonmulberry Silk Fibroin Scaffold Shows Superior Osteoconductivity Than Mulberry Silk Fibroin in Calvarial Bone Regeneration. *Adv Healthc Mater*. 2015;4(11). doi:10.1002/adhm.201500283
156. Xue Y, Kim HJ, Lee J, Liu Y, Hoffman T, Chen Y, et al. Co-Electrospun Silk Fibroin and Gelatin Methacryloyl Sheet Seeded with Mesenchymal Stem Cells for Tendon Regeneration. *Small*. 2022;18(21). doi:10.1002/sml.202107714
157. Zhou S, Wang Q, Yang W, Wang L, Wang J, You R, et al. Development of a bioactive silk fibroin bilayer scaffold for wound healing and scar inhibition. *Int J Biol Macromol*. 2024;255. doi:10.1016/j.ijbiomac.2023.128350
158. Li M, Tian W, Zhang Y, Song H, Yu Y, Chen X, et al. Enhanced Silk Fibroin/Sericin Composite Film: Preparation, Mechanical Properties and Mineralization Activity. *Polymers (Basel)*. 2022;14(12). doi:10.3390/polym14122466
159. Mushtaq A, Do KL, Wahab A, Yousaf M, Rahman A, Hussain H, et al. Silk Fibroin-Derived Smart Living Hydrogels for Regenerative Medicine and Organoid Engineering: Bioactive, Adaptive, and Clinically Translatable Platforms. *Gels*. 2025. doi:10.3390/gels11110908
160. Xing X, Han Y, Cheng H. Biomedical applications of chitosan/silk fibroin composites: A review. *International Journal of Biological Macromolecules*. 2023. doi:10.1016/j.ijbiomac.2023.124407
161. Chen Q, Wu K, Yao J, Shao Z, Chen X. Adhesive silk fibroin/magnesium composite films and their application for removable wound dressing. *Biomater Sci*. 2024;13(1). doi:10.1039/d4bm01411a
162. Zhao J, Zhang Z, Wang S, Sun X, Zhang X, Chen J, et al. Apatite-coated silk fibroin scaffolds to healing mandibular border defects in canines. *Bone*. 2009;45(3). doi:10.1016/j.bone.2009.05.026
163. Zhou K, Yuan T, Wang S, Hu F, Luo L, Chen L, et al. Beyond natural silk: Bioengineered silk fibroin for bone regeneration. *Materials Today Bio*. 2025. doi:10.1016/j.mtbio.2025.102014
164. Khosropanah MH, Ghofrani A, Vaghassloo MA, Zahir M, Bahrami A, Azimzadeh A, et al. Biomedical applications of Bombyx mori silk in skin regeneration and cutaneous wound healing. *Biomedical Materials (Bristol)*. 2025. doi:10.1088/1748-605X/adb552
165. Choi JH, Jeon H, Song JE, Oliveira JM, Reis RL, Khang G. Biofunctionalized lysophosphatidic acid/silk fibroin film for cornea endothelial cell regeneration. *Nanomaterials*. 2018;8(5). doi:10.3390/nano8050290
166. Wang L, Chen Z, Yan Y, He C, Li X. Fabrication of injectable hydrogels from silk fibroin and angiogenic peptides for vascular growth and tissue regeneration. *Chemical Engineering Journal*. 2021;418. doi:10.1016/j.cej.2021.129308
167. Cui X, Wang X, Liu S, Wang L, Li M, Zhang J, et al. Silk fibroin-based biomaterials for spinal cord



- injury repair: Recent advances and future prospects. *BMEMat.* 2025. doi:10.1002/bmm2.70026
168. Orash Mahmoud Salehi A, Nourbakhsh MS, Rafienia M, Baradaran-Rafii A, Heidari Keshel S. Corneal stromal regeneration by hybrid oriented poly (ϵ -caprolactone)/lyophilized silk fibroin electrospun scaffold. *Int J Biol Macromol.* 2020;161. doi:10.1016/j.ijbiomac.2020.06.045
169. Rama M, Vijayalakshmi U. Influence of silk fibroin on the preparation of nanofibrous scaffolds for the effective use in osteoregenerative applications. *J Drug Deliv Sci Technol.* 2021;61. doi:10.1016/j.jddst.2020.102182
170. Gao L, Li Y, Liu G, Lin X, Tan Y, Liu J, et al. Mechanical properties and biocompatibility characterization of 3D printed collagen type II/silk fibroin/hyaluronic acid scaffold. *J Biomater Sci Polym Ed.* 2025;36(5). doi:10.1080/09205063.2024.2411797
171. Karageorgiou V, Meinel L, Hofmann S, Malhotra A, Volloch V, Kaplan D. Bone morphogenetic protein-2 decorated silk fibroin films induce osteogenic differentiation of human bone marrow stromal cells. *J Biomed Mater Res A.* 2004;71(3). doi:10.1002/jbm.a.30186
172. Xiao W, Liu W, Sun J, Dan X, Wei D, Fan H. Ultrasonication and genipin cross-linking to prepare novel silk fibroin-gelatin composite hydrogel. *J Bioact Compat Polym.* 2012;27(4). doi:10.1177/0883911512448692
173. Vishwanath V, Pramanik K, Biswas A. Optimization and evaluation of silk fibroin-chitosan freeze-dried porous scaffolds for cartilage tissue engineering application. *J Biomater Sci Polym Ed.* 2016;27(7). doi:10.1080/09205063.2016.1148303
174. Chou KC, Chen CT, Cherng JH, Li MC, Wen CC, Hu SI, et al. Cutaneous regeneration mechanism of β -sheet silk fibroin in a rat burn wound healing model. *Polymers (Basel).* 2021;13(20). doi:10.3390/polym13203537
175. Sabarees G, Tamilarasi GP, Jayaraman R, Alagarsamy V, Solomon VR. Silk Fibroin Hydrogels: Cutting-Edge Developments and Future Directions. *Pharm Nanotechnol.* 2024;13. doi:10.2174/0122117385339249241102165029
176. Liu J, Xie X, Wang T, Chen H, Fu Y, Cheng X, et al. Promotion of Wound Healing Using Nanoporous Silk Fibroin Sponges. *ACS Appl Mater Interfaces.* 2023;15(10). doi:10.1021/acsami.2c20274
177. Chen X, Li Z, Ge XX, Qi X, Xiang Y, Shi Y, et al. Ferric Iron/Shikonin Nanoparticle-Embedded Hydrogels with Robust Adhesion and Healing Functions for Treating Oral Ulcers in Diabetes. *Advanced Science.* 2024;11(45). doi:10.1002/advs.202405463
178. Ghosh S, Shajahan F, Adhikari J, Bera AK, Ghosh A, Pati F. Visible Light Cross-Linked Methacrylated Silk Fibroin Enables Enhanced Osteogenic Response in Bioprinted Dual-Layer Guided Bone Regeneration Membrane. *ACS Appl Mater Interfaces.* 2025;17(16). doi:10.1021/acsami.4c22349
179. Zhang Y, Roohani I. Recent Advances in Silk Fibroin Derived from *Bombyx mori* for Regenerative Medicine. *J Funct Biomater.* 2025;17(1). doi:10.3390/jfb17010012
180. An J, Ma T, Wang Q, Zhang J, Santerre JP, Wang W, et al. Defining optimal electrospun membranes to enhance biological activities of human endometrial MSCs. *Front Bioeng Biotechnol.* 2025;13. doi:10.3389/fbioe.2025.1551791
181. Lee DH, Tripathy N, Shin JH, Song JE, Cha JG, Min KD, et al. Enhanced osteogenesis of β -tricalcium phosphate reinforced silk fibroin scaffold for bone tissue biofabrication. *Int J Biol Macromol.* 2017;95. doi:10.1016/j.ijbiomac.2016.11.002
182. Varma R, Aoki FG, Soon K, Karoubi G, Waddell TK. Optimal biomaterials for tracheal epithelial grafts: An in vitro systematic comparative analysis. *Acta Biomater.* 2018;81. doi:10.1016/j.actbio.2018.09.048
183. Dsouza MV, Dodamani S, Kurangi B, Kumbar V, Hussain M. Functionalized Nanoscaffolds for Drug Delivery Using Biomaterials: A Glimmer of Hope in Diabetic Wound Management. *Regenerative Engineering and Translational Medicine.* 2025. doi:10.1007/s40883-025-00415-y
184. Nakazawa Y. Structural analysis and application to biomaterials of the silk fibroins. *Kobunshi Ronbunshu.* 2010;67(8). doi:10.1295/koron.67.428



185. Wang G, Liu R, Ma H, Duan S, Yang G, Meng LX, et al. Elevator-Like Hollow Channels in Porous Scaffolds Accelerate Vascularized Bone Regeneration via NETs-Fibrin-Mediated Macrophage Recruitment. *Advanced Science*. 2025. doi:10.1002/advs.202515693
186. Singh BN, Pramanik K. Development of novel silk fibroin/polyvinyl alcohol/sol-gel bioactive glass composite matrix by modified layer by layer electrospinning method for bone tissue construct generation. *Biofabrication*. 2017;9(1). doi:10.1088/1758-5090/aa644f
187. Ma D, Wang Y, Dai W. Silk fibroin-based biomaterials for musculoskeletal tissue engineering. *Materials Science and Engineering C*. 2018. doi:10.1016/j.msec.2018.04.062
188. Mamdoohi M, Shafieian M, Hassannejad Z. A Triple-Layered Composite Scaffold of Silk Fibroin and Decellularized Amniotic Membrane for Bladder Tissue Engineering. *Macromol Biosci*. 2025;25(11). doi:10.1002/mabi.202500157
189. Choi JH, Kim DK, Song JE, Oliveira JM, Reis RL, Khang G. Silk Fibroin-Based Scaffold for Bone Tissue Engineering. In: *Advances in Experimental Medicine and Biology*. 2018. doi:10.1007/978-981-13-0947-2_20
190. Bi X, Mao Z, Zhang Y, Ren Z, Yang K, Yu C, et al. Endogenous dual-responsive and self-adaptive silk fibroin-based scaffold with enhancement of immunomodulation for skull regeneration. *Biomaterials*. 2025;320. doi:10.1016/j.biomaterials.2025.123261
191. Flanagan KE, Tien LW, Elia R, Wu J, Kaplan D. Development of a sutureless dural substitute from Bombyx mori silk fibroin. *J Biomed Mater Res B Appl Biomater*. 2015;103(3). doi:10.1002/jbm.b.33217
192. Ribeiro VP, Pina S, Gheduzzi S, Araújo AC, Reis RL, Oliveira JM. Hierarchical HRP-Crosslinked Silk Fibroin/ZnSr-TCP Scaffolds for Osteochondral Tissue Regeneration: Assessment of the Mechanical and Antibacterial Properties. *Front Mater*. 2020;7. doi:10.3389/fmats.2020.00049
193. Yang C, Li S, Huang X, Chen X, Shan H, Chen X, et al. Silk Fibroin Hydrogels Could Be Therapeutic Biomaterials for Neurological Diseases. *Oxidative Medicine and Cellular Longevity*. 2022. doi:10.1155/2022/2076680
194. Khademolqorani S, Banitaba SN, Kouhi M, Behrouznejad B. Harnessing the Osteogenic Potential of Novel Copper Modified Baghdadite Nanogalleries Integrated in Silk Fibroin Electrospun Scaffolds for Enhanced Bone Regeneration. *J Polym Environ*. 2025;33(7). doi:10.1007/s10924-025-03598-1
195. Vidya M, Rajagopal S. Silk Fibroin: A Promising Tool for Wound Healing and Skin Regeneration. *International Journal of Polymer Science*. 2021. doi:10.1155/2021/9069924
196. Yan S, Wang Q, Tariq Z, You R, Li X, Li M, et al. Facile preparation of bioactive silk fibroin/hyaluronic acid hydrogels. *Int J Biol Macromol*. 2018;118. doi:10.1016/j.ijbiomac.2018.06.138
197. Karimi F, Farbehi N, Ziaee F, Lau K, Monfared M, Kordanovski M, et al. Photocrosslinked Silk Fibroin Microgel Scaffolds for Biomedical Applications. *Adv Funct Mater*. 2024;34(29). doi:10.1002/adfm.202313354
198. Huh JT, Lee JU, Kim WJ, Yeo M, Kim GH. Preparation and characterization of gelatin/ α -TCP/SF biocomposite scaffold for bone tissue regeneration. *Int J Biol Macromol*. 2018;110. doi:10.1016/j.ijbiomac.2017.09.030
199. Hama R, Aytemiz D, Moseti KO, Kameda T, Nakazawa Y. Silk Fibroin Conjugated with Heparin Promotes Epithelialization and Wound Healing. *Polymers (Basel)*. 2022;14(17). doi:10.3390/polym14173582