



# Cathelicidin LL-37 Level in Leprosy Patients and Household Contacts - A Systematic Review

Nur Hasira Mustakim<sup>a</sup>, Widya Widita<sup>a</sup>, Asnawi Madjid<sup>a</sup>, Khairuddin Djawad<sup>a</sup>, Andi Nurhaerani Zainuddin<sup>a</sup>

<sup>a</sup>Department of Dermatology and Venereology, Faculty of Medicine, University of Hasanuddin, Makassar, Indonesia, 90245

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## KEYWORDS

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## ABSTRACT:

Cathelicidin LL-37 is a multifunctional antimicrobial peptide central to innate immunity. Its dysregulated expression in leprosy patients and household contacts suggests a significant role in pathogenesis and potential as a prognostic biomarker. This systematic review followed PRISMA guidelines, searching PubMed, Cochrane Library, and Google Scholar for studies published between 2010–2025. Inclusion criteria focused on human observational research assessing LL-37 at protein or gene levels in serum, skin, or blood. From 55,212 records, four studies were analyzed. Matzner et al. (2010) identified significantly lower serum LL-37 in untreated leprosy patients. Conversely, Argentina et al. (2023a, 2023b) reported elevated LL-37 protein and *CAMP* gene expression in skin lesions, with intermediate levels found in household contacts. Oliveira et al. (2022) observed reduced *CAMP* and vitamin D receptor expression in peripheral blood across both patients and contacts, correlating with inflammatory cytokine profiles that persisted post-therapy. The evidence reveals a compartmentalized paradox: systemic suppression of LL-37 alongside localized overexpression in lesional skin. Systemic downregulation may be linked to impaired vitamin D–*CAMP* pathways, while local upregulation reflects innate activation against bacillary load. Intermediate levels in contacts suggest immunological priming from exposure. The vitamin D–cathelicidin axis emerges as a potential therapeutic target and a valuable biomarker for disease activity and risk stratification in endemic regions.

## 1. Introduction

Cathelicidins are a class of multifunctional antimicrobial peptides that play a central role in the innate immune defense system. This peptide is widely recognized for its broad-spectrum antimicrobial activity, demonstrating the ability to combat a wide array of pathogens, including those resistant to conventional antibiotics. Its action involves disrupting microbial membranes, immunomodulating, and enhancing host defense pathways, underscoring its importance in maintaining immune surveillance and resilience against infectious agents<sup>1</sup>. Beyond its antimicrobial action, cathelicidin has also been implicated in the pathophysiology of several inflammatory and infectious diseases. Elevated levels of this peptide have been reported in conditions such as pneumonitis and Inflammatory Bowel Disease (IBD), where it is thought to contribute to disease progression as well as modulation of inflammatory responses<sup>1,2</sup>.

In the context of leprosy, a chronic infectious disease caused by *Mycobacterium leprae*, cathelicidin expression has demonstrated distinct alterations. Gene expression analyses have revealed that cathelicidin is significantly elevated in skin lesions of leprosy patients compared to their normal skin, and also shows higher levels in both household contacts and patients when contrasted with healthy, non-exposed individuals<sup>3,4</sup>. This pattern of increased expression suggests that cathelicidin may serve not only as a marker of host immune activation but also as a potential biomarker for prognosis and treatment stratification. The ability to differentiate expression profiles between patients, contacts, and healthy controls has strong implications, especially in endemic areas where close household exposure is a major risk factor. Epidemiological data indicate that the incidence of leprosy among household contacts is approximately 1.4 cases per 1,000 person-years, with a cumulative finding of 192 out of 9,024 contacts



developing leprosy over a 33-year follow-up period<sup>5</sup>. Such data highlight the necessity of identifying predictive biomarkers that can distinguish between individuals at risk of disease progression and those who remain resistant despite exposure.

Given the persistent global burden of leprosy, particularly in endemic regions such as Brazil and parts of Asia, the development of reliable biomarkers like cathelicidin is of increasing significance. Traditional diagnostic methods, including slit-skin smears and histopathological examinations, often have limitations in sensitivity and cannot always predict disease evolution or exposure-related risk. Therefore, the prospect of cathelicidin serving as a prognostic tool could facilitate earlier diagnosis, more targeted treatment, and improved monitoring of household contacts. Its potential incorporation into clinical practice may allow for stratified therapeutic approaches, ultimately supporting the goal of reducing transmission and disability associated with the disease<sup>6,7</sup>.

## 2. Materials and Method

This systematic review aims to quantify and synthesize the differences in Cathelicidin LL-37 level among leprosy patients and their household contacts, while adhering to PRISMA guideline. The PICO for the following study are:

- **Population:** Humans with leprosy (any Ridley-Jopling class) and/or **household/close contacts**; healthy controls eligible. (Terminology around “contacts” is heterogeneous; we’ll accept study-defined contacts and capture definitions.)
- **Index/Exposure:** Cathelicidin/LL-37 measured in any matrix (serum/plasma, skin biopsy, PBMCs; mRNA, protein).
- **Comparator:** Healthy or non-exposed controls; between-group contrasts (patients vs contacts, contacts vs healthy).
- **Outcomes:** Primary—level/expression of LL-37 (or CAMP/hCAP18) as reported (pg/mL, AU, fold-change,  $\Delta$ Ct). Secondary—associations with clinical form (PB/MB), reactions (ENL/Type 1), treatment status, vitamin D/VDR.
- **Design:** Observational (cross-sectional, case-control, cohort); interventional if LL-37

measured at baseline or outcome. No date/language limits.

## 3. Search Strategy

Article search will be conducted on three databases (Pubmed, Cochrane Library and Google Scholar) using search queries tailored for those individual databases. Search process will include Randomized Clinical Trial, Cohort Study and any research articles while excluding any case reports and review articles.

**Pubmed:** ("Cathelicidins"[MeSH] OR cathelicidin\*[tiab] OR "LL-37"[tiab] OR LL37[tiab] OR hCAP18[tiab]) AND ("Leprosy"[MeSH] OR leprosy[tiab] OR "Hansen's disease"[tiab] OR "Hansen disease"[tiab] OR "Mycobacterium leprae"[tiab])

**Cochrane Library:** (leprosy OR Hansen OR "Mycobacterium leprae"):ti,ab,kw AND (cathelicidin OR "LL-37" OR LL37 OR hCAP18):ti,ab,kw

**Google Scholar:** "leprosy" OR "Hansen's disease" OR "Mycobacterium leprae" (LL 37 OR "cathelicidin" OR hCAP18)

## 4. Selection and data extraction

Authors independently reviewed titles, abstracts, and full texts. Four studies ; Matzner et al. (2011, Yemen), two studies by Argentina et al. (2023a,b, Indonesia), and Oliveira et al. (2022, Brazil), were included. Data extracted included author, year, country, study design, sample size, biospecimens, assay methods, comparator groups, and main outcomes. Household contacts were defined as individuals living in the same residence as leprosy patients.

## 5. Data synthesis and analysis

Study characteristics and results were narratively summarized and presented in tabular form. Quantitative data were extracted as means  $\pm$  SD, medians with interquartile ranges, or fold-change values where reported. Because of heterogeneity in study designs, populations, and measurement matrices (serum vs skin), no pooled meta-analysis was attempted. Instead, results were synthesized descriptively, highlighting systemic versus local expression patterns and the position of household contacts relative to patients and healthy groups.



6. Result

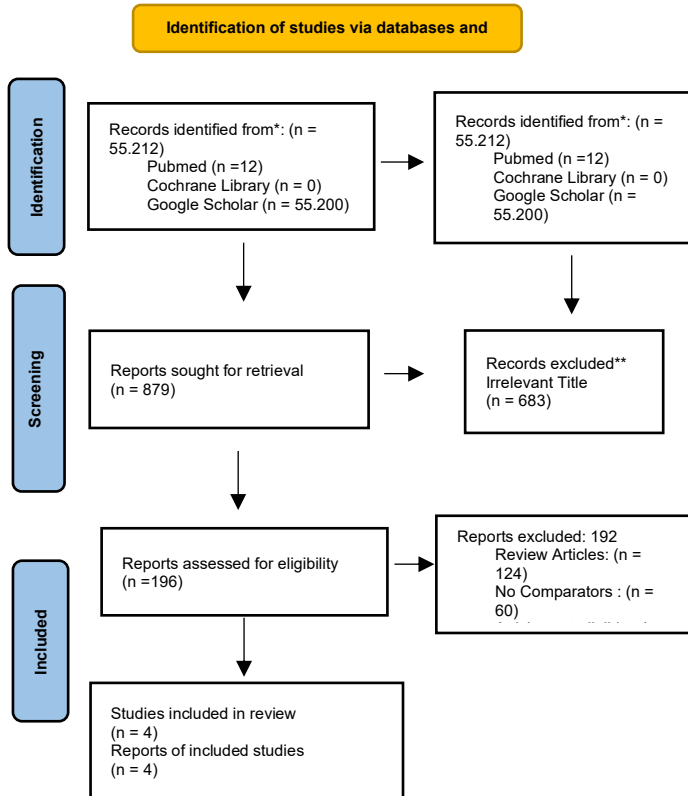


Figure 1 PRISMA Guideline for Article selection process

Table 1 Study Synthesis

Author, Year	Population	Sample / Method	Comparator	Key Findings
Matzner et al., 2010 (Acta Trop)	29 leprosy patients (14 untreated, 15 treated), 19 healthy controls (Yemen)	Serum, ELISA (LL-37), Vitamin D immunoassay	Patients vs untreated vs treated	Serum LL-37 significantly lower in leprosy patients vs healthy ( $p < 0.001$ ). Untreated patients had lowest levels (median 48 ng/mL), treated higher (median 98 ng/mL) but still < controls (median 160 ng/mL). Vitamin D3 levels not different. Suggests LL-37 suppressed in active disease.
Argentina et al., 2023 (Acta Med Acad)	18 leprosy patients, 18 household contacts, 18 cathelicidin healthy	Skin scrapings, ELISA (cathelicidin gene) and HBD-3	Patients vs contacts vs healthy	LL-37 protein: Leprosy > contacts > healthy. Mean $\pm$ SD: patients 256.8 $\pm$ 22.9 pg/mL; contacts 25.9 $\pm$ 2.7; healthy 1.4 $\pm$ 0.1 ( $p < 0.0001$ ). Demonstrates intermediate elevation in HHCs.
Argentina et al., 2023 (Clin Cosmet)	18 leprosy patients, 18 household contacts, 18 healthy	Skin swabs, qPCR for CAMP (cathelicidin gene) and HBD-3	Patients vs contacts vs healthy	CAMP expression: markedly $\uparrow$ in leprosy lesion skin (median 38.72 fold), intermediate in contacts (9.8), baseline in healthy (1.0). Normal skin of patients lower

Author, Year	Population	Sample / Method	Comparator	Key Findings
Invest				(0.48). Strongly significant ( $p < 0.0001$ ).
Derm)				HBD-3 also upregulated.
Oliveira et al., 2022 (Microbes Infect)	34 leprosy patients (PB & MB, untreated + 6mo MDT), 25 household contacts, 18 healthy (Brazil)	Peripheral blood, RT-qPCR for VDR & CAMP: serum cathelicidin & cytokines (ELISA & bead array)	Patients vs contacts vs healthy; pre vs post MDT	Reduced CAMP & VDR expression in patients and HHCs vs healthy (log10FC < -1 to -2). Expression correlated with IL-2, IFN- $\gamma$ , IL-17F. Path analysis: VDR/CAMP regulate inflammatory cytokine cascades. Persistent inflammatory profile even after 6mo MDT. Suggests impaired vitamin D-cathelicidin axis in disease.

7. Study Description

Matzner et al. (2010)

This study, conducted in Yemen, assessed serum LL-37 and vitamin D3 concentrations in 29 leprosy patients (14 untreated, 15 treated) and 19 healthy controls. Using ELISA, the authors found that serum LL-37 levels were markedly lower in both untreated and treated leprosy patients compared to controls. The lowest levels were detected in newly diagnosed, untreated patients, while treated patients showed partial restoration of LL-37 but remained below healthy levels. No significant differences in serum vitamin D3 were observed across groups. The study concluded that cathelicidin suppression may be linked to active *M. leprae* infection, independent of vitamin D status.

Argentina et al. (2023a) – Acta Medica Academica

This cross-sectional study in Indonesia compared cathelicidin protein levels in skin scrapings from 18 leprosy patients, 18 household contacts, and 18 healthy controls. Cathelicidin levels were quantified using ELISA, showing a gradient pattern: highest in patients (mean 256.8  $\pm$  22.9 pg/mL), intermediate in household contacts (25.9  $\pm$  2.7 pg/mL), and lowest in healthy individuals (1.4  $\pm$  0.1 pg/mL). Statistical analysis confirmed significant differences between groups ( $p < 0.0001$ ). The findings suggest that household contacts exhibit partial innate immune activation, reflecting their close exposure to *M. leprae*.



## Argentina et al. (2023b) – Clinical, Cosmetic and Investigational Dermatology

In this related study, also based in Indonesia, 18 leprosy patients, 18 household contacts, and 18 healthy controls were examined for gene expression of cathelicidin (CAMP) and human beta-defensin-3 (HBD-3). Skin swabs from lesional and non-lesional sites were analyzed using quantitative PCR. Results demonstrated a marked increase in CAMP expression in leprosy lesions (median 38.72 fold), intermediate expression in household contacts (9.8 fold), and baseline in healthy controls (1.0). Normal skin from patients had much lower expression (0.48 fold). HBD-3 followed a similar pattern. These results highlight local upregulation of antimicrobial peptide genes in response to bacillary exposure, in both patients and contacts.

## Oliveira et al. (2022)

This Brazilian study analyzed 34 leprosy patients (14 paucibacillary, 20 multibacillary), 25 household contacts, and 18 healthy controls. Peripheral blood samples were assessed for gene expression of vitamin D receptor (VDR) and CAMP using RT-qPCR, alongside serum vitamin D, cathelicidin, and cytokine levels. Both CAMP and VDR expression were significantly reduced in patients and contacts compared to healthy controls ( $\log_{10}FC < -1$  to  $-2$ ), and strong correlations were observed with inflammatory cytokines (IL-2, IFN- $\gamma$ , IL-17F). Importantly, these alterations persisted even after six months of multidrug therapy. Path analysis suggested that VDR–CAMP interactions modulate the inflammatory response to *M. leprae*. The authors concluded that impaired vitamin D–cathelicidin signaling contributes to chronic inflammation in leprosy.

## 8. Discussion

Leprosy, otherwise known as Hansen's Disease, is a chronic infectious disease that is caused by *Mycobacterium leprae* that primarily affects the skin and peripheral nerves. This bacteria is a slow-growing bacterium of which the infection will eventually leads to nerve damage and skin lesions<sup>8</sup>. This disease causes a range of symptoms which includes skin injuries, muscle weakness, numbness and in more advance cases may lead to disfiguring mutilations and disabilities such as blindness and paralysis<sup>6</sup>.

Diagnosis of this condition often involves clinical examination for cardinal signs that is then supported by laboratory tests such as slit-skin smear and histopathology<sup>9</sup>. Due to its inflammatory nature, biomarkers can be used to accurately diagnose the condition. Bioinformatics analysis showed 10 gene markers which includes ITK, CD48, IL2RG, CCR5, FGR, JAK3, STAT1, LCK, PTPRC, and CXCR4 which are considered to be an active immune system pathway for Chemokine signalling and T-Cell receptor pathway<sup>10</sup>.

Another form of biomarker that can and should be considered in the treatment and diagnostic of leprosy are peptides. These molecules play a significant role in the pathogenesis and treatment strategy for leprosy. For example, a specific peptide has been designed to target phenolic glycolipid-1 (PGL-1) of *Mycobacterium leprae*, which is crucial for the bacterium's entry into Schwann cells. This peptide, derived from the laminin subunit alpha-2 (LAMA2), was shown to potentially block the interaction between PGL-1 and Schwann cells, thereby inhibiting the initial stages of infection. The peptide's stability and preferential binding site were confirmed through molecular dynamics simulations, suggesting its potential as a therapeutic agent to prevent neural damage in leprosy<sup>11</sup>.

Peptide that is specifically studied in this review is Cathelicidin LL-37 which exhibits antimicrobial activity against a wide range of pathogens, including bacteria, viruses and fungi by targeting bacterial membrane lipids and other structures, disrupting their integrity and leading to cell death<sup>12</sup>. The earliest specific study that was found in this study is done by Matzner et al (2010), which showed that circulating LL-37 concentrations in serum are significantly reduced in leprosy patients compared to healthy individuals, with untreated cases demonstrating the lowest levels. This finding suggests a systemic suppression of cathelicidin in active infection. Importantly, vitamin D levels were not significantly different between patients and controls, raising the possibility of vitamin D–independent mechanisms contributing to LL-37 downregulation in leprosy<sup>13</sup>.

In contrast, more recent studies evaluating skin-derived cathelicidin revealed increased expression in lesional tissue. Argentina et al. (2023) demonstrated that LL-37 protein levels were markedly higher in the skin of leprosy patients than in household contacts and healthy



individuals, with contacts showing intermediate concentrations. This supports the concept that continuous exposure to *M. leprae* can prime the innate immune system of household contacts, leading to partial upregulation of antimicrobial peptides. A complementary gene expression study by the same group confirmed that CAMP mRNA was elevated in leprosy lesions and, to a lesser extent, in household contacts, relative to healthy skin<sup>3,4</sup>. Together, these findings highlight a divergence between systemic and local expression, where blood levels appear reduced while tissue-specific expression is enhanced in response to bacillary burden.

The mechanistic insights from Oliveira et al. (2022) further elucidate this paradox. Their study showed that both CAMP and vitamin D receptor (VDR) gene expression were significantly reduced in the peripheral blood of patients and household contacts compared to healthy controls. This downregulation correlated with inflammatory cytokine profiles, including IL-2, IFN- $\gamma$ , and IL-17F, and persisted despite six months of multidrug therapy. These results suggest that the vitamin D–CAMP axis is impaired in leprosy and may underpin the systemic reduction in LL-37 observed in serum. At the same time, persistent inflammatory signaling could explain the increased cathelicidin detected locally in skin tissue<sup>14</sup>.

In terms of its function, it has a dual function in pro-inflammation and immuno-response, which are important in fighting bacterial infections, but, under certain condition it has been observed to be able to inhibit inflammatory process<sup>15</sup>. Beyond leprosy, LL-37 has been studied in other conditions such as sepsis where it shows a release of antimicrobial ectosome in neutrophils, reducing bacterial load and improving survival of patients with sepsis<sup>16</sup>.

Furthermore, it has also been studied in its role in other inflammatory diseases, such as Type 2 Diabetes and Chronic Lung illness, highlighting its multifunctional nature in immune regulation and disease outcome<sup>17-20</sup>.

Having established a potential prognostic modality for Leprosy, it is worth noting that there are other biomarkers that can be used in identifying leprosy, such as ITK, CD48, IL2RG, CCR5, FGR, JAK3, STAT1, LCK, PTPRC, and CXCR4, which are active in immune

system pathways such as Chemokine signaling and T cell receptor signaling<sup>21</sup>. Tumour Necrosis Factor (TNF) and Interleukin 10 (IL-10) is also considered to an accurate biomarker for classifying leprosy patients and household contacts<sup>22</sup>. PCR-based detection can also be used in detecting the main cause of Leprosy, which is the *Mycobacterium Leprae*, in which the PCR can be used to detect the bacterium's DNA<sup>23</sup>.

Elevated levels of IgA antibodies, which occurred due to reaction of *Mycobacterium Leprae*-specific antigens such as NDO-HSA, LID-1, and NDO-LID, can be used as biomarkers for leprosy detection, aiding in the diagnosis and classification of the disease, especially in multibacillary patients and household contacts<sup>24-27</sup>.

Treating leprosy requires a multi-drug treatment, which is the primary treatment. The standard multi-drug treatment involved is a regiment of rifampicin, dapsone and clofazimine, which is effective for both paucibacillary and multibacillary type of leprosy<sup>28,29</sup>. The main challenge in treating leprosy is one of the more drug therapy problem which is drug resistance, relapses and the need for new therapeutic agents, thus it is important, going forward, to improve and overcome this persisting problem<sup>30</sup>.

## 9. Conclusion

Cathelicidin LL-37 is dysregulated in leprosy, showing reduced systemic levels but enhanced local expression in lesions and contacts. These consistent differences across compartments underline its dual role in immune defense and inflammation. LL-37 and its regulatory pathways may serve as valuable biomarkers for exposure and disease activity, and future studies should further explore their diagnostic and therapeutic potential.

## References

1. Krawiec P, Pac-Kożuchowska E. Cathelicidin – a novel potential marker of pediatric inflammatory bowel disease. *Journal of Inflammation Research*. 2021;14:163–74.
2. Lemieszek MK, Golec M, Zwoliński J, Dutkiewicz J, Milanowski J. Cathelicidin influence on pathological activation of Wnt pathway in murine model of hypersensitivity pneumonitis. *Annals of Agricultural and Environmental Medicine*. 2022;29(3):358–64.



3. Argentina F, Suwarsa O, Gunawan H, Berbudi A. Gene Expression of Human Beta-Defensin-3 and Cathelicidin in the Skin of Leprosy Patients, Household Contacts, and Healthy Individuals from Indonesia. *Clinical, Cosmetic and Investigational Dermatology*. 2023;16:1485–92.
4. Argentina F, Suwarsa O, Gunawan H, Berbudi A. A Comparison of Cathelicidin Levels in the Skin of Leprosy Patients and Their Household Contacts. *Acta Medica Academica*. 2023 Dec 1;52(3):195–200.
5. Hacker MA, Sales AM, Duppre NC, Sarno EN, Moraes MO. Leprosy incidence and risk estimates in a 33-year contact cohort of leprosy patients. *Scientific Reports*. 2021 Dec 1;11(1).
6. Slater KB. A Current Perspective on Leprosy (Hansen’s Disease). In: *Vaccines for Neglected Pathogens: Strategies, Achievements and Challenges: Focus on Leprosy, Leishmaniasis, Melioidosis and Tuberculosis*. Springer International Publishing; 2023. p. 29–46.
7. Wang Y, Xiao D, Wu M, Qing L, Yang T, Xiao P, et al. Epidemiological Characteristics and Factors Associated with Cure of Leprosy in Chongqing, China, from 1949 to 2019. *American Journal of Tropical Medicine and Hygiene*. 2023 Jan 1;108(1):165–73.
8. Berto Pucca M. “Leprosy Overview: Pathophysiology, Immune Responses, and Epidemiology in Brazil.” *Biomedical Journal of Scientific & Technical Research*. 2023 Feb 10;48(4).
9. Bathula S, Khurana A, Singh I. Diagnosis of Leprosy: Current Updates and Future Directions. *Indian Journal of Postgraduate Dermatology*. 2023 Feb 7;1:13–23.
10. Zhou Q, Shi P, Shi W dong, Gao J, Wu Y chen, Wan J, et al. Identification of potential biomarkers of leprosy: A study based on GEO datasets. *PLoS ONE*. 2024 May 1;19(5 May).
11. Arenas NE, Pieffet G, Rocha-Roa C, Guerrero MI. Design of a specific peptide against phenolic glycolipid-1 from *Mycobacterium leprae* and its implications in leprosy bacilli entry. *Memorias do Instituto Oswaldo Cruz*. 2022;117.
12. Zeth K, Sancho-Vaello E. Structural plasticity of ll-37 indicates elaborate functional adaptation mechanisms to bacterial target structures. Vol. 22, *International Journal of Molecular Sciences*. MDPI; 2021.
13. Matzner M, al Samie AR, Winkler HM, Nemeth J, Grasnek A, Indra A, et al. Low serum levels of cathelicidin LL-37 in leprosy. *Acta Tropica*. 2011 Jan;117(1):56–9.
14. Grossi de Oliveira AL, Chaves AT, Cardoso MS, Pinheiro GRG, Antunes DE, Grossi MA de F, et al. Reduced vitamin D receptor (VDR) and cathelicidin antimicrobial peptide (CAMP) gene expression contribute to the maintenance of inflammatory immune response in leprosy patients. *Microbes and Infection*. 2022 Sep 1;24(6–7).
15. Yang B, Good D, Mosaiab T, Liu W, Ni G, Kaur J, et al. Significance of LL-37 on Immunomodulation and Disease Outcome. Vol. 2020, *BioMed Research International*. Hindawi Limited; 2020.
16. Kumagai Y, Murakami T, Kuwahara-Arai T, Iba T, Reich J, Nagaoka I. Antimicrobial peptide LL-37 ameliorates a murine sepsis model via the induction of microvesicle release from neutrophils. *Innate Immunity*. 2020 Oct 1;26(7):565–79.
17. Armiento V, Hille K, Naltsas D, Lin JS, Barron AE, Kapurniotu A. The Human Host-Defense Peptide Cathelicidin LL-37 is a Nanomolar Inhibitor of Amyloid Self-Assembly of Islet Amyloid Polypeptide (IAPP). *Angewandte Chemie - International Edition*. 2020 Jul 27;59(31):12837–41.
18. Golec M, Lemieszek MK, Dutkiewicz J, Milanowski J, Barteit S. A Scoping Analysis of Cathelicidin in Response to Organic Dust Exposure and Related Chronic Lung Illnesses. Vol. 23, *International Journal of Molecular Sciences*. MDPI; 2022.
19. Suzuki K, Ohkuma M, Someya A, Mita T, Nagaoka I. Human Cathelicidin Peptide LL-37 Induces Cell Death in Autophagy-Dysfunctional Endothelial Cells. *The Journal of Immunology*. 2022 May 1;208(9):2163–72.
20. Bryzek D, Golda A, Budziaszek J, Kowalczyk D, Wong A, Bielecka E, et al. Citrullination-resistant ll-



- 37 is a potent antimicrobial agent in the inflammatory environment high in arginine deiminase activity. *International Journal of Molecular Sciences*. 2020 Dec 1;21(23):1–15.
21. Hooij A van, Fat EMTK, Eeden SJ van den, Wilson L, da Silva MB, Salgado CG, et al. Field-friendly serological tests for determination of *M. leprae*-specific antibodies. In: *Development of immunodiagnostic tests for leprosy: from biomarker discovery to application in endemic areas* [Internet]. 2021. Available from: <https://hdl.handle.net/1887/3240171>
22. Marçal PHF, de Souza MLM, Gama RS, de Oliveira LBP, Gomes MDS, do Amaral LR, et al. Algorithm Design for a Cytokine Release Assay of Antigen-Specific in Vitro Stimuli of Circulating Leukocytes to Classify Leprosy Patients and Household Contacts. *Open Forum Infectious Diseases*. 2022 Mar 1;9(3).
23. Manta FS de N, Leal-Calvo T, Moreira SJM, Marques BLC, Ribeiro-Alves M, Rosa PS, et al. Ultra-sensitive detection of mycobacterium leprae: DNA extraction and PCR assays. *PLoS Neglected Tropical Diseases*. 2020 May 1;14(5):1–15.
24. Silva KK de P e., Oliveira EE de, Elias CMM, Pereira IE, Pinheiro RO, Sarno EN, et al. Serum IgA Antibodies Specific to *M. leprae* Antigens as Biomarkers for Leprosy Detection and Household Contact Tracking. *Frontiers in Medicine*. 2021 Aug 10;8.
25. Gama RS, Leite LA, Colombo LT, Fraga LA de O. Prospects for new leprosy diagnostic tools, a narrative review considering elisa and PCR assays. *Revista da Sociedade Brasileira de Medicina Tropical*. 2020;53:1–7.
26. Longoni SS, Beltrame A, Prato M, Spencer JS, Bergamaschi N, Clapasson A, et al. ELISA Test Based on the Phenolic Glycolipid-I (PGL-I) of *Mycobacterium leprae*: A Reality of a Laboratory from a Non-Endemic Country. *Pathogens*. 2022 Aug 1;11(8).
27. Leal-Calvo T, Avanzi C, Mendes MA, Benjak A, Busso P, Pinheiro RO, et al. A new paradigm for leprosy diagnosis based on host gene expression Insights from leprosy lesions transcriptomics. *bioRxiv* [Internet]. 2021 Jul 30; Available from: <http://biorxiv.org/lookup/doi/10.1101/2021.07.30.454441>
28. Susanto PM, Esti PK, Komarasari E. Effectivity of Uniform Multidrug Therapy on the Success of Paucibacillary and Multibacillary Leprosy Treatment. *Nepal Journal of Dermatology, Venereology & Leprology*. 2022 Sep 30;20(2):16–23.
29. Denisa Alinda M, Geani S, Indira Agusni R, Haryo Kusumaputra B, Rizky Reza N, Rosita Sigit Prakoeswa C, et al. Diagnosis and Management of Leprosy. *Periodical of Dermatology and Venereology*. 2020;32(2).
30. Divya C, Raj K, Aditya S. Therapy of Leprosy-Present Strategies and Recent Trends with Immunotherapy. *Journal of Dermatology Research and Therapy*. 2020 Dec 31;6(2).