

Extraction and Determination of Trace Copper (II) Using Octadecyl Silica Membrane Disks Modified 1-(2-Pyridyl Azo) 2-Naphtol(Pan) in Water Samples and Paraffin-Embedded Tissues from Liver Loggerhead Turtles Specimens by FAAS

A. Moghimi^{1*}, M. Shabanzadeh²

¹Department of Chemistry, Varamin (Pishva) Branch, Islamic Azad University, Varamin, Iran

²Department of Chemistry, Damghan Branch, Islamic Azad University, Damghan, Iran

Abstract: A simple and reproducible method for the rapid extraction and determination of trace amounts of copper(II) ions using octadecyl-bonded silica membrane disks modified by 1-(2-Pyridyl Azo)2-Naphtol(PAN) and Atomic Absorption Spectrometry was presented. The method was based on complex formation on the surface of the ENVI-18 DISK™ disks followed by stripping of the retained species by minimum amounts of appropriate organic solvents. The elution was efficient and quantitative. The effect of potential interfering ions, pH, ligand amount, stripping solvent, and sample flow rate were also investigated. Under the optimal experimental conditions, the break-through volume was found to be about 1000mL providing a preconcentration factor of 500. In the present study, we reported the application of preconcentration techniques still continues increasingly for trace metal determinations by flame atomic absorption spectrometry (FAAS) for quantification of Cu in Formalin-fixed paraffin-embedded (FFPE) tissues from Liver loggerhead turtles. The maximum capacity of the disks was found to be $389 \pm 4 \mu\text{g}$ for Cu^{2+} . The limit of detection of the proposed method was 5ng per 1000mL. The method was applied to the extraction and recovery of copper in different water samples.

Keywords: Copper (II), SPE, Octadecyl silica disks, AAS, 1-(2-Pyridyl Azo) 2-Naphtol (PAN) Formalin-fixed paraffin-embedded (FFPE)

INTRODUCTION

Copper at trace concentrations acts as both a micronutrient and a toxicant in marine and fresh water systems [1- 8]. This element is needed by plants at only very low levels and is toxic at higher levels. At these levels, copper can be bound to the cell membrane and hinder the transport process through the cell wall. Copper at nearly 40ng mL^{-1} is required for normal metabolism of many living organisms [9, 10]. On the other hand, copper is an important element in many industries. Thus, the development of new methods for selective separation, concentration and determination of it in

sub-micro levels in different industrial, medicinal and environmental samples is of continuing interest. The determination of copper is usually carried out by flame and graphite furnace atomic absorption spectrometry (AAS) [11, 12] as well as spectrometric methods [13, 14]. However, due to the presence of copper in medicinal and environmental samples at low levels, its separation from other elements presents and also the use of a preconcentration step prior to its determination is usually necessary.

Corresponding author: A. Moghimi, Department of Chemistry, Varamin(Pishva) Branch, Islamic Azad University, Varamin, Iran E-mail: alimoghimi@iauvaramin.ac.ir

Different methods, especially Liquid- Liquid extraction of copper in the presence of various classical [15-19] and macrocyclic[20, 21] co-extractant ligands has attracted considerable attention. However, the use of classical extraction methods for this purpose is usually time-consuming, labor-intensive and requires large amounts of high purity solvents for extraction. Nevertheless, several other techniques for the preconcentration and separation of copper have been proposed including liquid chromatography [22] supercritical fluid extraction [23], flotation [24], aggregate film formation [25], liquid membrane [26], column adsorption of pyrocatechol violet-copper complexes on activated carbon [27], ion pairing [28], ion pairing [29], preconcentration with yeast [30], and solid phase extraction using C₁₈ cartridges and disks [31-33, 56].

Solid phase extraction (SPE) or liquid-solid extraction is popular and growing techniques that are used to sample preparation for analysis. It is an attractive alternative for classical liquid-liquid extraction methods that reduce solvent usage and exposure, disposal costs and extraction time for Sample separation and concentration purposed [34-36]. In recent years, the octadecyl-bonded silica SPE disks have been utilized for the extraction and separation of different organic compounds from environmental matrices [37-40]. Moreover, the SPE disks modified by suitable ligands are successfully used for selective extraction and concentration of metal ions [41-42].

In a recent series of papers, [43-45] we have described the application of metal-DNA conjugates to nucleic acid sequence determination with catalytic signal amplification; the assay relies on the esterase activity of a DNA-linked Cu complex. For optimization of the system and exploration of structure- activity relationships, a sensitive probe would be useful, which allows straightforward detection of esterase activity of ligated Cu²⁺ in low

concentration. The tabulated structures reveal that atoms (NNOO or NNSS) are proper completing agents for the determination of lead. In the present report, we wish to describe a proper concentrative method for assessment of trace levels of lead in different water samples. To the best of our knowledge, octadecyl silica membrane disks modified by the introduced symmetrical Schiff's base (PAN) (Fig 1) have not been used for lead isolation and preconcentration, previously (Table 1) of this work was the development of a rapid, efficient and highly sensitive method for selective extraction and concentration of ultra trace amounts of Cu²⁺ ions from aqueous media using octadecyl silica membrane disks modified by 1-(2-Pyridyl Azo)2-Naphtol(PAN)and AAS determination.

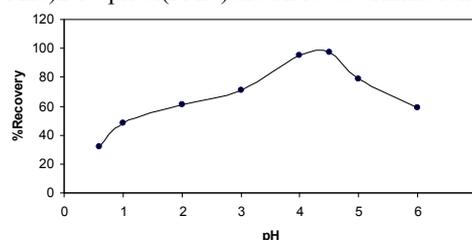


Fig 1. Influence of sample pH and dissolving solvent of PAN on the percentage recovery of Cu(II)

Table 1. The effect of the different Schiff's base molecular structures on their selectivity toward different metal traces by using C₁₈ octadecyl silica bonded phase sorbent

Schiff's base	Ion(s) of interest	Ref.
	Pb ⁺²	58
	Pb ⁺²	59
	Cu ⁺²	60
	Cu ⁺² & Pb ⁺²	61
	Cu ⁺² & Pb ⁺²	62

The optimized method was applied to Cu^{2+} determinations in different natural waters. The second aim of this study was the selection of an appropriate method for the analysis of FFPE tissue was based on present work with atomic absorption spectrophotometric determination of Cu.

EXPERIMENT

Reagents

All acids were of the highest purity available from Merck and were used as received. Methanol and Chloroform were of HPLC grade from Merck. Analytical grade nitrate salts of lithium, sodium, potassium, magnesium, calcium, strontium, barium, zinc, cadmium, lead, nickel, cobalt(II), and copper(II) were of the highest purity. Ultra pure organic solvents were obtained from E.Merck, Darmstat, Germany, and high purity double distilled deionized water was used throughout the experiments.

The stock standard solution of Cu^{2+} was prepared by dissolving 0.1000g of the copper powder in 10mL concentrated nitric acid and diluted to 1000mL with water in a calibrated flask. Working solutions were prepared by appropriate dilution of the stock solution.

Apparatus

Determination of Cu^{2+} contents in working samples were carried out by a Varian spectra A.200 model atomic absorption spectrometer equipped with a high intensity hollow cathode lamp(HI-HCL) according to the recommendations of the manufacturers. These characteristics are tabulated in (Table 2).

Table 2. The operational conditions of flame for determination of copper

Slit width	0.7 nm
Operation current of HI-HCL	15 mA
Resonance fine	324.8nm
Type of background correction	Deuterium lamp
Type of flame	Air/acetylene
Air flow	7.0 mL.min ⁻¹
Acetylene flow	1.7 mL.min ⁻¹

Solid phase extractions were carried out by glassy membrane disks, ENVI-18DISK™ 47mm diameter ×0.6 mm thickness containing octadecyl silica bonded phase (30 μm particles, 70 Å° pore size) obtained from Supelco in conjunction with a standard Millipore 47 mm filtration apparatus equipped with a vacuum pump. The pH measurements were carried out by an ATC pH meter (EDT instruments, GP 353).

Sample extraction

Extraction was performed with glassy membrane disks, ENVI-18 DISK™ 47mm diameter ×0.6 mm thickness containing octadecyl silica bonded phase (30 μm particles, 70 Å° pore size) from Supelco. The disks were used in conjunctions with a standard Millipore 47mm filtration apparatus connected to water aspirator [46].

1) *Sample Treatment:* The water samples were filtered through 45μm nylon filters. Sampling vessels were polyethylene bottles soaked in 1 mol.L⁻¹ HNO₃ overnight and rinsed twice with deionized water. The analysis must be done within 2 days of sample collection to limit the risk of interconversion of copper (II).Then; 5mL of methanol was added to a 90mL portion of each before analysis. The surface of the ENVI-18 DISK™ disks is not modified with PAN and therefore could not retain Cu^{2+} ions properly. Instead, 10 mg of PAN was dissolved in an appropriate volume of an organic solvent (5mL) miscible with water. The most suitable solvent under the experimental conditions was acetone. The PAN solution was added to aqueous solution of Cu^{2+} and the mixture was stirred gently.

2) *Disk cleaning and conditioning:* A disk was placed in the apparatus and was washed with 10mL of methanol to remove all contaminants arising from the manufacturing process and the

environment. Then, the disk was dried by passing air through it for several minutes. To insure optimal extraction of the analytes of interest, the disk was again washed with 10mL of methanol, immediately followed by 10mL of water, without letting the surface of the disk dry. This step pre-wets the disk surface prior to extraction. Improper performance of this step causes slow flow – rate and poor analyte recoveries. It is important to avoid any air contact with the surface of the disk before the addition of the sample.

3) *Sample addition:* After complete homogenization, accurate volumes of the sample solutions (100mL portions) were transferred to the top reservoir of the disk apparatus. At the same time, the solution was drawn through the disk by applying a mild vacuum. Application of vacuum was continued until the disk was completely dry (about 5 minute).

4) *Analyte elution:* In order to elute the analyte selectively, exactly 5 mL of acidified solvents 0.1M HCl in methanol was passed through the disk and collected into a 5.0 mL volumetric flask under the extraction funnel. It was found that ultra pure alcoholic organic solvents were the best eluting agents. The concentration of copper (II) in the eluates were then determined by FAAS using an external calibration graph .

2.7 Analysis of Sample paraffin-embedded tissues from Liver loggerhead turtles specimens

Selected areas from fresh frozen tissues from Liver loggerhead turtles specimens were sliced in three pieces (numbered as 1, 2 and 3) of approximately 10mm×5mm×2mm each. Sets of pieces of set 1 (controls), were placed into a vacuum chamber at 50 °C overnight to dry (until a constant weight was obtained), and the sets 2 and 3 were subjected to the standard 10% buffered formalin fixation and paraffin embedding [55]. histological process using

a tissue processor (Tissue-Tek VIP, Sakura Finetek USA Inc., Torrance, CA). After the paraffin embedding process, tissues were subsequently excised from the blocks with a titanium knife and deparaffinized in xylene at 55 °C for 1 h in the tissue processor (the set 2), or with hexane at 20 °C for 1 week with frequent changes of the solvent in handling-based procedure (the set 3). Xylene was of a grade routinely used for the FFPE process and hexane was of “Optima” grade (Fisher Scientific). Upon deparaffinization, the tissue samples were dried in a vacuum chamber until constant weight was obtained. Each dried sample (of the sets 1–3) was divided into three portions (5–10 mg each) to be further analyzed as triplicates.

RESULTS AND DISCUSSION

Evaluation of the role of the ligand

Some preliminary experiments were performed for investigation of absence or presence of PAN on the quantitative extraction of copper (II). It was concluded that the membrane disk itself does not show any tendency for the retention of copper (II), but introduction of 100mL portions of aqueous copper (II) samples containing 10µg of copper (II) and 10mg of PAN leads to satisfactory its retention (Table 3). The latter case is most probably attributed to the existence of a considerable interaction between copper (II) and the PAN. It should be mentioned that formation of stable complexes between copper (II) and PAN at pH=2 is probably due to an ion pair formation mechanism. However, at pH higher than 2 the retention and percentage recovery of copper (II) are negligible .

Table 3. The effect of presence of PAN on extraction percent of copper(II)^a

PAN	pH	Extraction percent of copper(II)
Absence	2-6	0.03(6.7) ^b
Presence	2-6	98.6(2.7) to 65(2.8)

^a Initial samples contained 10µg of copper(II) in 100mL of water.

^b Values in parentheses are RSDs based on five individual replicate analyses

Choice of eluent

In order to select the most appropriate eluent for the quantitative stripping of the retained copper (II) on the disks, 5mL of various non organic (each containing 10% V.V⁻¹ methanol) and different organic solvents were tested. The results were tabulated in Table2. As can be seen, the best eluting solvents were found to be 5mL of methanol or ethanol, resulting in quantitative elution of copper (II) from the disk. It should be emphasized that presence of methanol in any kind of employed solvents helps to better the contact of eluent with hydrophobic surface of the disk .

The effect of the pH

The pH of the sample solutions were adjusted to different values between 2-9 by addition of hydrochloric acid or a suitable buffer such as sodium acetate-acetic acid or sodium dihydrogen phosphate- disodium hydrogen phosphate, and then solutions passed through the disks.

Eventually, the metal ions were stripped by pure methanol or ethanol solutions followed by flame atomic absorption determination of the eluted copper (II). Then, percentage recovery at various pH values was determined (Fig .1). According to the results shown in Fig.1 up to pH 4.0-4.5, complete recoveries were obtained. However, at higher pH values, percentage recovery decreases. This is due to fact that in an acidic solution the protonation of PAN occurs and there is a weak tendency for retention between Cu (II) and PAN, whereas at higher values (pH>5), Cu (II) reacts with hydroxide ions to produce Cu (OH) 2. Therefore, sodium acetate-acetic acid buffer with pH=4.5 was used for the preconcentration step. Other solvents used for dissolving PAN were methanol and ethanol. The influences of these solvents on the recoveries as a function of pH were compared and shown in Fig. 1. Meanwhile, other organic solvents were not tested because of their

restricted solubility and formation of two phases with aqueous solutions and incompatibility with flame higher pH values (>7) were not tested because of the possibility of the hydrolysis of octadecyl silica in the disks[47]. Cu(II) ions can be retained quantitatively by the modified membrane disk through the pH range from 4.0 to 4.5 However, at lower pH (< 4.0), nitrogen atoms of the PAN could be protonated and the stability of complex was reduced.

Effect amount of counter anion

In order to investigate the effect of counter ion on the recovery Cu²⁺ ions by the modified disks, different counter anions were tested Table 4, it is immediately obvious that the nature of the counter anion strongly influences the retention of copper ions by the disk. The results revealed that the PAN behaves as a neutral ionophore in the pH range 4.0-4.5 [49, 50]. So that the copper ions were retained as ion pair complexes by the membrane disks. As seen, acetate ion is the most efficient counter anion for the SPE of Cu (II) ions. The influence of the concentration of sodium acetate ion was investigated on copper recovery, and the results are shown in Table 4. As seen, the percent recovery of Cu²⁺ increased with the acetate concentration until a reagent concentration of about 0.1 M was reached, beyond which the recovery remained quantitative.

Table 4. Percent recovery of copper from the modified membrane disk in the presence of 0.01 M of different counter anions ^a

counter anion	%Recovery
Cl ⁻	26.5
Br ⁻	24.7
ClO ₄ ⁻	35.6
SCN ⁻	45.6
Picrate	74.9
Acetate	98.5

^a Initial samples contained 10µg of copper(II) in 100mL of water

Moreover, acted ion acted as a suitable buffering agent, while it effectively contributed to the ion-pair formation; thus, in the SPE experiments, there was no need for the addition of any buffer solution.

Effect of eluent acidity

Cu (II) complexes is eluted quantitatively only with acidified methanol (> 0.1 M in HCl or HNO₃).

Hence, acidified methanol (0.1 M HCl in methanol) was used to strip the Cu (II) sorbed on membrane disk in the presence of 1-(2-Pyridyl Azo) 2-Naphtol (PAN) chelate (Table 5).

Table 5. Effect of different eluting solvents on Percentage recovery of copper(II) adsorbed on the disk ^a

Stripping solution	% Recovery		
	2ml	5ml	10ml
Methanol	86(2.6) ^b	98.5(2.6)	98.0(2.7)
Acidified methanol ^c	57(2.3)	87.2(2.2)	83.9(1.7)
Ammoniacal methanol ^d	57 (2.5)	87.5(2.6)	86.9(2.2)
Ethanol	89.1(1.7)	100(1.5)	99.8(2.3)
Acetonitril	36(4.8)	46(5.5)	69(3.7)
Formic acid(1M)10% V.V ⁻¹ mthanol	55(1.2)	68(2.0)	71(2.2)
Hydrochloric acid(1M)10% V.V-1 mthanol	59(1.9)	92.0(2.4)	91(2.9)
Hydrochloric acid(1M)10% V.V ⁻¹ mthanol	53(2.5)	85 (2.6)	93(1.7)
Nitric acid(2M)10% V.V ⁻¹ mthanol	53(1.9)	86(2.2)	87(2.0)
Nitric acid(1M)10% V.V ⁻¹ mthanol	64(2.5)	85(2.3)	89(1.7)
Ethanol	83(2.8)	95.5(2.0)	97.7(2.9)

^a Initial samples contained 10 µg of each copper in 100 mL water

^b Values in parentheses are RSDs based on five individual replicate analysis

^c Acidified solvents obtained by addition of 0.1M HCl

^d Ammoniacal solvents obtained by addition of 0.1M NH₃

The influence of flow-rate

One of the most important parameters affecting solid phase extraction is the speed of the process. Hence, the effect of flow-rates on extraction efficiencies was investigated. It was found that in the range of 10-100 mL.min⁻¹, the retention of Cu (II) was not considerably affected by the sample solutions flow-rates and led to reproducible and satisfactory results (Fig 2). Thus, the flow-rate was maintained at 89mL.min⁻¹ throughout the experiment .

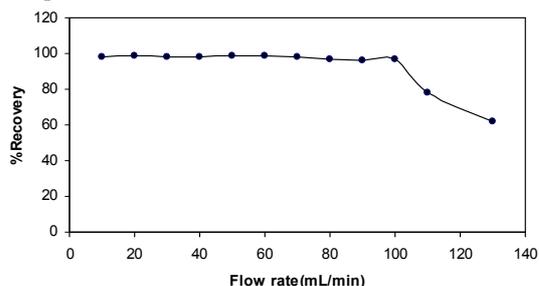


Fig 2. The effect of the flow-rate on extraction percent of Cu(II)

Quantity of the PAN

The optimum amount of PAN for the quantitative extraction of Cu (II) was also investigated by adding various amounts of it to solution (between 2-20 mg). The results are listed in Table 6. The experimental results revealed that the extraction of Cu (II) was quantitative using a sample solution containing more than 10 mg PAN. Hence, subsequent extractions were performed with 15mg of PAN.

Table 6. In fluence of the PAN amount on the recovery of Cu(II) ions ^a

PAN amount (mg)	Recovery(%) of Cu(II)
2	36(2.2) ^b
5	48(2.9)
8	89(2.7)
10	97.8(2.9)
15	98.9(2.6)
20	98.7(2.5)

^a Initial samples contained 10 µg of each copper in 100 mL water

^b Values in parentheses are RSDs based on five individual replicate analysis

Disk efficiency

Undoubtedly, one of the major parameters affecting in the SPE determinations is the efficiency of the used membrane disks. However, to the best of our knowledge this case has not been discussed elsewhere in similar reports. Under the optimum experimental conditions, it was found out that each ENV-18 DISK™ disk could perform at least 14 replicate analyses if organic eluting solvents were used. On the other hand, acidic, eluents practically decrease the number of time a disk could be used to 10 replicates. These observations are represented in Fig 3.

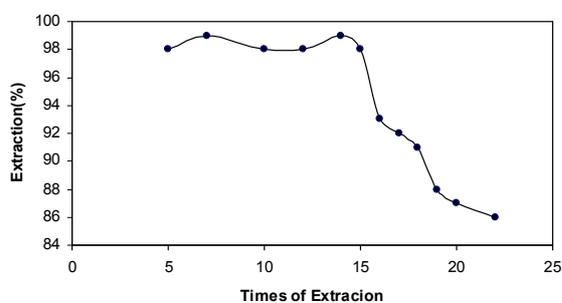


Fig 3. Influence of eluent(5mL of methanol)type on disk efficiency

Analytical Performance

When solutions of 10µg copper in 10, 50, 100, 500, 1000, 2000, 2500 and 3000mL solutions under optimal experimental conditions were passed through the disks, the Cu (II) was quantitatively retained in all cases. Thus, the breakthrough volume for the method must be greater than 2500mL, providing a concentration factor of >500. The limit of detection (LOD) of the method for the determination of Cu (II) was studied under the optimal experimental conditions. The LOD based on 3σ of the blank (5mL of methanol) is 5 ng per 1000mL.

The capacity of modified disks (5mg PAN) was determined by passing 50mL portions of sample solutions containing 8mg of copper and 0.1M sodium acetate-acetic acid buffer with pH 4.0-4.5, followed by the determination of the retained metal

ions in the eluting solution using AAS. The maximal capacity of the disk obtained from three replicate measurements was 389±4µg of Cu²⁺ on the disk .

In order to investigate the selective separation and determination of Cu²⁺ ions from its binary mixtures with various metal ions, an aliquot of aqueous solutions (50mL) containing 10µg Cu²⁺ and mg amounts of other cations was taken and the recommended procedure was followed. The results are summarized in Table 7. The results show that the copper (II) ions in binary mixtures were retained almost completely by the modified disk, even in the presence of up to about 100mg of various ions. Meanwhile, retention of other cations by the disk is very low and they can be separated effectively from the Cu²⁺ ion. Its is interesting to note that, in other experiments, we found that in the presence of high enough concentrations NH₂OH.HCl as a suitable reducing agent (> 0.5M) 48. No retention of the resulting single charge Cu⁺ ion can be occurred by the modified membrane disk.

Table 7. Separation of copper from binary mixtures ^a

Diverse ion	Amounts taken(mg)	% Found	%Recovery of Cu ²⁺ ion
Na ⁺	92.4	1.15(2.4) ^b	98.7(1.8)
K ⁺	92.5	1.32(2.3)	98.5(2.2)
Mg ²⁺	14.5	0.7(1.2)	98.9(1.7)
Ca ²⁺	26.3	2.25(3.0)	98.5(1.8)
Sr ²⁺	2.45	2.85(2.1)	98.4(2.0)
Ba ²⁺	2.66	3.16(2.1)	98.3(2.3)
Mn ²⁺	2.66	1.75(2.2)	96.3(2.8)
Co ²⁺	2.13	1.4(2.3)	99.1(2.9)
Ni ²⁺	1.67	2.0(2.4)	98.5(2.4)
Zn ²⁺	2.78	1.97(2.1)	98.6(2.2)
Cd ²⁺	2.58	1.92(0)	98.2(2.8)
Pb ²⁺	0.54	2.7(1.9)	97(2.7)
Hg ²⁺	0.43	2.81(2.1)	97.7(2.8)
Ag ⁺	2.67	3.45(2.9)	96.6(2.9)
Cr ³⁺	1.76	2.92(2.30)	97.3(2.4)
UO ²⁺	2.86	2.8(2.1)	98.3(2.7)

^a Initial samples contained 10µg Cu²⁺ and different amounts of various ions in 100 mL water(0.1 M acetate ion)

^b Values in parentheses are RSDs based on five individual replicate analysis

Analysis of a real biological sample

To assess the applicability of the method to real samples, it was applied to the extraction and determination of copper from different water samples. Tap water (Tehran, taken after 10 min operation of the tap), rain water (Tehran, 26 January, 2006), Snow water (Saveh, 6 February, 2006) and Sea water (taken from Caspian Sea, near the Mahmoud-Abad shore) samples were analyzed (Table 8). As can be seen from Table 4 the added copper ions can be quantitatively recovered from the water samples used. As it is seen, the recovered copper ion revealed that the results are quite reliable and are in satisfactory agreement with those obtained by ICPAES.

Development of a methodology for the determination of Cd in FFPE tissue was performed in a number of steps to optimize the major factors affecting the precision of the analysis (Table 8).

Table 8. Recovery of copper added to 1000mL of different water samples (containing 0.1M acetate at pH= 4.0-4.5)

Sample	Cu ²⁺ added (µg)	Cu ²⁺ determined (ng.mL ⁻¹)	ICP-AES
Tap water	0.0	1.74(1.8) ^a	ND
	10.0	11.98(2.2)	11.7
Snow water	0.0	4.86(2.3)	ND
	10.0	14.94(2.0)	14.7
Rain water	0.0	2.66(2.3)	ND
	10.0	12.76(2.4)	12.3
Sea Water	0.0	12.67(2.3)	12.5
	10.0	22.95(2.0)	23.1
Development of a methodology for the determination of Cd in FFPE tissue	0.0	-	
	10.0	10.07(0.9)	

^a Values in parentheses are %RSDs based on five individual replicate analysis

^b Not detected

CONCLUSION

Results presented in this work demonstrate well the tremendous possibilities offered by the solid phase extraction of trace amounts of Cu(II) in water

samples using Octadecyl Silica membrane disks modified by 1-(2-Pyridyl Azo)2-Naphtol(PAN) and its determination by FAAS. The method developed was simple, reliable, and precise for determining copper in water. Also, the proposed method was free of interference compared to conventional procedures to determine copper [51-55, 57]. The method can be successfully applied to the separation and determination of copper in binary mixtures.

ACKNOWLEDGEMENTS

The author wish to thank the Chemistry Department of Varamin branch Islamic Azad University for financial support.

REFERENCES

1. H.J.M.Bowen.1979 .Environmental Chemistry of the Elements, Academic Press, New York, 132-135 .
2. L.E.Brand, W.G. Sunda,R.R.L. Guillard.1986 J.Exp.Mar.Biol.Ecol.96:225.
3. H.H. Taylor, J.M.Anstiss.1999, Mar. Freshwat. Res.50:907.
4. F.M.M.Morel, R.J.M. Hudson, N.M.Price, 1991 Limnol.Oceanogr.36:1742.
5. A.S.Gordan, 1992 Mar.Chem.38:1.
6. J.W.Moffett, L.E.Brand, P.L. Croot, K.A.Barbeau1997 ,Limnol.Oceanogr. 42:789.
7. P.L.Croot,J.W.Moffett, L.E.Brand,2000 Limnol.Oceanogr.45:619.
8. M.Wood, H.K.Wang, 1983 Environ.Sci.Technol.17:582A.
9. N.N.Greenwood, A.Eamshow, 1984 Chemistry of Elements, Pergamon Press, New York.
10. C.A.Burtis, E.R.Ashwood,Tiets 1999 Textbook of Clinical Chemistry, third ed., Macmillan,New York.
11. B. Wetz, 1985 Atomic Absorption Spectroscopy, VCH, Amsterdam.

12. A.D.Eaton, L.S.Clesceri, A.E.Greenberg, 1995 Standard Methods for the examination of water and waste water ,19 th ed ,American Public Health Association, Washington,DC .
13. F.J.Welcher, E.Boschmann, 1979 Organic Reagents for Copper, Krieger Huntington, New York.
14. Z.Marczenko, 1986 Separation and Spectrophotometric Determination of Elements, Ellis Horwood, London.
15. O.P.Bharagava, 1969 Talanta 16:743.
16. A.A. Schilt, W.C. Hoyle, 1964 Anal. Chem. 41:344.
17. L.G.Borchart, J.P. Butler, 1957 Anal. Chem. 29:414.
18. R. Chaisuksant, W.P. Ayuthaya, K. Grudpan, 2000 Talanta 53:579.
19. D. Kara, M. Alkan, 2002 Microchem. J. 71: 29.
20. K.Saito, S.Murakami, A. Muromatsu, E.Sekido, 1994 Anal.Chim. Acta 294:329.
21. K. Ikeda, S. Abe, 1998 Anl.Chim. Acta 363: 165.
22. S. Igarashi, N. Ide, Y.Takagai, 2000 Anal.Chim. Acta 424:263.
23. J. Liu, W.Wang, G. Li, 2001 Talanta 53:1149.
24. A.N.Anthemidis, G.A. Zachariadis, J.A. Stratis, 2001 Talanta 54:935.
25. D. Zenedelovska, G. Pavlovska, K. Cundeva, T.Stafilov, 2001 Talanta 54:139.
26. M.Endo, K.Suzuki, S.Abe, 1998 Anal. Chim. Acta 364:13.
27. M.E. Campderros, A. Acosta, J. Marchese, 1998 Talanta 47:19.
28. I.Narin, M. Soylak, L.Elic, M.Dogan, 2000 Talanta 52:1041.
29. Y.Akama, M.Ito, S.Tanaka, 2000 Talanta 52:645.
30. K.Ohta, H.Tanahasi, T. Suzuki, S.Kaneco, 2001, Talanta 53:715.
31. V.Cuculic, M.Mlakar, M.Branica, Anal. Chim. Acta 339:(1997)181.
32. A.Moghimi, M.S.Tehrani, S.Waqif Husain, 2006 Material Science Research India 3(1a) :27.
33. M.S.Tehrani, A.Moghimi, S.Waqif Husain, 2005 Material Science Research India 3(2) :135.
34. E.M.Thurman, M.S.Mills, 1998 Solid-Phase Extraction, Principles and Practice, Wiley, New York.
35. J.Pawliszyn, 1997 Solid-Phase Microextraction, Theory and Practice, Wiley-VCH, New York.,
36. R.M. Izatt, J.S. Bradshaw, R.L. Bruening, 1996 Pure Appl. Chem. 68:1237.
37. D.F.Hagen, C.G.Markell, G.A. Schmitt, 1990 Anal.Chim. Acta 236:157.
38. C.J.Krueger, J.A. Fild, 1995 Anal.Chem. 67:3363.
39. K.Z.Taylor, D.S.Waddell, E.J.Reiner, 1995 Anal. Chem. 67:1186.
40. Y.Yamini, M.Ashraf-Khorassani, 1994 J.High Resolut. Chromatogr.17:634.
41. M.Shamsipur, A.R.Ghiasvand, Y.Yamini, 1999 Anal.Chem. 71:4892.
42. M.Shamsipur, A.R.Ghiasvand, H. Sharghi, 2001 Int. J. Environ. Anal.Chem. 82:23.
43. Brunner, J.; Mokhir, A.; Kramer, R. 2003 J.Am.Chem.Soc. 125:12410.
44. Zelder, F.H.; Brunner, J.; Kramer, R. 2004 Chem. Commun., 902.
45. Boll, I.; Kramer, R.; Brunner, J.; Mokhir, A. J.Am.Chem. Soc. 27(2005)7849.
46. Hagen, D.F.; Mrkell, C.G.; Schmitt, G.A.; Blevins, D.D.; Anal. Chim. Acta 236(1990) 157 .
47. Yamini, Y.; Alizadeh, N.; Shamsipur, M.; Anal. Chim. Acta 69(1997) 355.
48. Petit de Pena, Y.; Gallego M.; Valcarcel, M.; 1995 Anal. Chem. 67:2524 .
49. Alizadeh, N.; Naeimi, H.; Sharghi, H.; Samsipur, Polish, M.; 1999 Polish J.Chem.73:915.

50. Moghimi A, Poursharifi M.J. 2011, Australian Journal of Basic and Applied Sciences, 5(6): 979-986 ,
51. Choi,Y.S.;Choi,H.S. 2003 Bull.Korean Chem. Soc.24:222.
52. Matoso,E.;Kubota,L.T.;Cadore,S. 2003 STalanta 60:1105.
53. Purachat, B.; Liawruangrath,S.;Sooksamiti,P.;Rattanaphani,S.;Buddhasukh, D.2001 Anal.Sci. 17:443.
54. Ensafi,A.A.;Abbasi,S.;Rahimi Mansour,H.;Mohammad pour Baltork, I. 2001.Anal.Sci. 17:609.
55. Saber Tehrani,M.; Rastegar,F.; Parchehbaf, A.;Rezvani,Z,2005.Chinese Journal of Chemistry 23:1437.
56. Moghimi A, R. Ghiasi, 2008 J. Korean Chem. Soc., 52: 4.
57. Moghimi A., 2007; Chinese Journal of Chemistry 12:1842-1847.
58. Hashemi, O. R.; Kargar-Razi, M.; Raoufi, F.; Moghimi, A.; Aghabozorg, H.; Ganjali, M. R. 2001 Microchem. J. 69: 1
59. Hashemi, O. R.; Raoufi, F.; Ganjali, M. R.; Moghimi, A.; Kargar-Razi, M.; Aghabozorg, H.; Shamsipur, M. 2000,Anal. Sci. 16: 1221.
60. Ganjali,M R ; Pourjavid, M. R.; Hajiagha Babaei, L.;Niasari, M. S. 2004.Quim Nova 27: 213.
61. Ganjali,M. R.; Pourjavid, M. R.; Hajiagha Babaei, L.;Niasari, M. S. 2004. Ann. Chim. (Rome) 94: 47.
62. Ganjali,M R.; Basiripour, F.; Niasari, M. S. Can. 2002 J. Anal. Sci. Spect. 47: 41.